

# Package ‘jmv’

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**Description** A suite of common statistical methods such as descriptives, t-tests, ANOVAs, regression, correlation matrices, proportion tests, contingency tables, and factor analysis. This package is also useable from the ‘jamovi’ statistical spreadsheet (see <<https://www.jamovi.org>> for more information).

**License** GPL (>= 2)

**Depends** R (>= 3.2)

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## R topics documented:

ancova . . . . .	2
ANOVA . . . . .	5
anovaNP . . . . .	7

anovaOneW . . . . .	9
anovaRM . . . . .	11
anovaRMNP . . . . .	14
bugs . . . . .	15
cfa . . . . .	16
contTables . . . . .	19
contTablesPaired . . . . .	22
corrMatrix . . . . .	23
descriptives . . . . .	26
efa . . . . .	29
linReg . . . . .	31
logLinear . . . . .	34
logRegBin . . . . .	38
logRegMulti . . . . .	41
logRegOrd . . . . .	44
mancova . . . . .	47
pca . . . . .	48
propTest2 . . . . .	50
propTestN . . . . .	52
reliability . . . . .	53
ttestIS . . . . .	55
ttestOneS . . . . .	57
ttestPS . . . . .	60

**Index****63**

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ancovaANCOVA

---

**Description**

The Analysis of Covariance (ANCOVA) is used to explore the relationship between a continuous dependent variable, one or more categorical explanatory variables, and one or more continuous explanatory variables (or covariates). It is essentially the same analysis as ANOVA, but with the addition of covariates.

**Usage**

```
ancova(
  data,
  dep,
  factors = NULL,
  covs = NULL,
  effectSize = NULL,
  modelTest = FALSE,
  modelTerms = NULL,
  ss = "3",
  homo = FALSE,
```

```

norm = FALSE,
qq = FALSE,
contrasts = NULL,
postHoc = NULL,
postHocCorr = list("tukey"),
postHocES = list(),
emMeans = list(list()),
emmPlots = TRUE,
emmPlotData = FALSE,
emmPlotError = "ci",
emmTables = FALSE,
emmWeights = TRUE,
ciWidthEmm = 95,
formula
)

```

## Arguments

data	the data as a data frame
dep	the dependent variable from data, variable must be numeric (not necessary when providing a formula, see examples)
factors	the explanatory factors in data (not necessary when providing a formula, see examples)
covs	the explanatory covariates (not necessary when providing a formula, see examples)
effectSize	one or more of 'eta', 'partEta', or 'omega'; use eta <sup>2</sup> , partial eta <sup>2</sup> , and omega <sup>2</sup> effect sizes, respectively
modelTest	TRUE or FALSE (default); perform an overall model test
modelTerms	a formula describing the terms to go into the model (not necessary when providing a formula, see examples)
ss	'1', '2' or '3' (default), the sum of squares to use
homo	TRUE or FALSE (default), perform homogeneity tests
norm	TRUE or FALSE (default), perform Shapiro-Wilk tests of normality
qq	TRUE or FALSE (default), provide a Q-Q plot of residuals
contrasts	a list of lists specifying the factor and type of contrast to use, one of 'deviation', 'simple', 'difference', 'helmert', 'repeated' or 'polynomial'
postHoc	a formula containing the terms to perform post-hoc tests on (see the examples)
postHocCorr	one or more of 'none', 'tukey', 'scheffe', 'bonf', or 'holm'; provide no, Tukey, Scheffe, Bonferroni, and Holm Post Hoc corrections respectively
postHocES	a possible value of 'd'; provide cohen's d measure of effect size for the post-hoc tests
emMeans	a formula containing the terms to estimate marginal means for (see the examples)
emmPlots	TRUE (default) or FALSE, provide estimated marginal means plots

emmPlotData	TRUE or FALSE (default), plot the data on top of the marginal means
emmPlotError	'none', 'ci' (default), or 'se'. Use no error bars, use confidence intervals, or use standard errors on the marginal mean plots, respectively
emmTables	TRUE or FALSE (default), provide estimated marginal means tables
emmWeights	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency
ciWidthEmm	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
formula	(optional) the formula to use, see the examples

## Value

A results object containing:

results\$main	a table of ANCOVA results
results\$model	The underlying aov object
results\$assump\$homo	a table of homogeneity tests
results\$assump\$norm	a table of normality tests
results\$assump\$qq	a q-q plot
results\$contrasts	an array of contrasts tables
results\$postHoc	an array of post-hoc tables
results\$emm	an array of the estimated marginal means plots + tables

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$main$asDF
```

```
as.data.frame(results$main)
```

## Examples

```
data('ToothGrowth')
ancova(formula = len ~ supp + dose, data = ToothGrowth)

#
# ANCOVA
#
# ANCOVA
#
# -----
#          Sum of Squares   df   Mean Square    F     p
# -----
#    supp              205     1      205.4   11.4   0.001
#    dose              2224     1     2224.3  124.0  < .001
#    Residuals         1023    57      17.9
# 
#
ancova(
```

```
formula = len ~ supp + dose,  
data = ToothGrowth,  
postHoc = ~ supp,  
emMeans = ~ supp)
```

---

ANOVA

ANOVA

---

## Description

The Analysis of Variance (ANOVA) is used to explore the relationship between a continuous dependent variable, and one or more categorical explanatory variables.

## Usage

```
ANOVA(  
  data,  
  dep,  
  factors = NULL,  
  effectSize = NULL,  
  modelTest = FALSE,  
  modelTerms = NULL,  
  ss = "3",  
  homo = FALSE,  
  norm = FALSE,  
  qq = FALSE,  
  contrasts = NULL,  
  postHoc = NULL,  
  postHocCorr = list("tukey"),  
  postHocES = list(),  
  emMeans = list(list()),  
  emmPlots = TRUE,  
  emmPlotData = FALSE,  
  emmPlotError = "ci",  
  emmTables = FALSE,  
  emmWeights = TRUE,  
  ciWidthEmm = 95,  
  formula  
)
```

## Arguments

- |      |   |
|------|---|
| data | the data as a data frame  |
| dep  | the dependent variable from data, variable must be numeric (not necessary when providing a formula, see examples) |

<code>factors</code>	the explanatory factors in data (not necessary when providing a formula, see examples)
<code>effectSize</code>	one or more of 'eta', 'partEta', or 'omega'; use eta <sup>2</sup> , partial eta <sup>2</sup> , and omega <sup>2</sup> effect sizes, respectively
<code>modelTest</code>	TRUE or FALSE (default); perform an overall model test
<code>modelTerms</code>	a formula describing the terms to go into the model (not necessary when providing a formula, see examples)
<code>ss</code>	'1', '2' or '3' (default), the sum of squares to use
<code>homo</code>	TRUE or FALSE (default), perform homogeneity tests
<code>norm</code>	TRUE or FALSE (default), perform Shapiro-Wilk tests of normality
<code>qq</code>	TRUE or FALSE (default), provide a Q-Q plot of residuals
<code>contrasts</code>	a list of lists specifying the factor and type of contrast to use, one of 'deviation', 'simple', 'difference', 'helmert', 'repeated' or 'polynomial'
<code>postHoc</code>	a formula containing the terms to perform post-hoc tests on (see the examples)
<code>postHocCorr</code>	one or more of 'none', 'tukey', 'scheffe', 'bonf', or 'holm'; provide no, Tukey, Scheffe, Bonferroni, and Holm Post Hoc corrections respectively
<code>postHocES</code>	a possible value of 'd'; provide cohen's d measure of effect size for the post-hoc tests
<code>emmMeans</code>	a formula containing the terms to estimate marginal means for (see the examples)
<code>emmPlots</code>	TRUE (default) or FALSE, provide estimated marginal means plots
<code>emmPlotData</code>	TRUE or FALSE (default), plot the data on top of the marginal means
<code>emmPlotError</code>	'none', 'ci' (default), or 'se'. Use no error bars, use confidence intervals, or use standard errors on the marginal mean plots, respectively
<code>emmTables</code>	TRUE or FALSE (default), provide estimated marginal means tables
<code>emmWeights</code>	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency
<code>ciWidthEmm</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
<code>formula</code>	(optional) the formula to use, see the examples

## Details

ANOVA assumes that the residuals are normally distributed, and that the variances of all groups are equal. If one is unwilling to assume that the variances are equal, then a Welch's test can be used instead (However, the Welch's test does not support more than one explanatory factor). Alternatively, if one is unwilling to assume that the data is normally distributed, a non-parametric approach (such as Kruskal-Wallis) can be used.

## Value

A results object containing:

results\$main	a table of ANOVA results
results\$model	The underlying aov object
results\$assump\$homo	a table of homogeneity tests
results\$assump\$norm	a table of normality tests
results\$assump\$qq	a q-q plot
results\$contrasts	an array of contrasts tables
results\$postHoc	an array of post-hoc tables
results\$emm	an array of the estimated marginal means plots + tables

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$main$asDF
as.data.frame(results$main)
```

## Examples

```
data('ToothGrowth')
ANOVA(formula = len ~ dose * supp, data = ToothGrowth)

#
# ANOVA
#
# ANOVA
#
# -----
#          Sum of Squares   df   Mean Square    F     p
# -----
#   dose           2426    2      1213.2  92.00 < .001
#   supp           205     1       205.4  15.57 < .001
#   dose:supp     108     2        54.2   4.11  0.022
#   Residuals     712    54        13.2
# -----
# 

ANOVA(
  formula = len ~ dose * supp,
  data = ToothGrowth,
  emMeans = ~ supp + dose:supp, # est. marginal means for supp and dose:supp
  emmPlots = TRUE,             # produce plots of those marginal means
  emmTables = TRUE)            # produce tables of those marginal means
```

## Description

The Kruskal-Wallis test is used to explore the relationship between a continuous dependent variable, and a categorical explanatory variable. It is analogous to ANOVA, but with the advantage of being non-parametric and having fewer assumptions. However, it has the limitation that it can only test a single explanatory variable at a time.

## Usage

```
anovaNP(data, deps, group, es = FALSE, pairs = FALSE, formula)
```

## Arguments

data	the data as a data frame
deps	a string naming the dependent variable in data
group	a string naming the grouping or independent variable in data
es	TRUE or FALSE (default), provide effect-sizes
pairs	TRUE or FALSE (default), perform pairwise comparisons
formula	(optional) the formula to use, see the examples

## Value

A results object containing:

results\$table	a table of the test results
results\$comparisons	an array of pairwise comparison tables

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$table$asDF
as.data.frame(results$table)
```

## Examples

```
data('ToothGrowth')

anovaNP(formula = len ~ dose, data=ToothGrowth)

#
# ONE-WAY ANOVA (NON-PARAMETRIC)
#
# Kruskal-Wallis
# -----
#      X²      df     p
# -----
#      len    40.7     2   < .001
# -----
```

---

anovaOneW*One-Way ANOVA*

---

## Description

The Analysis of Variance (ANOVA) is used to explore the relationship between a continuous dependent variable, and one or more categorical explanatory variables. This 'One-Way ANOVA' is a simplified version of the 'normal' ANOVA, allowing only a single explanatory factor, however also providing a Welch's ANOVA. The Welch's ANOVA has the advantage that it need not assume that the variances of all groups are equal.

## Usage

```
anovaOneW(
  data,
  deps,
  group,
  welchs = TRUE,
  fishers = FALSE,
  miss = "perAnalysis",
  desc = FALSE,
  descPlot = FALSE,
  norm = FALSE,
  qq = FALSE,
  eqv = FALSE,
  phMethod = "none",
  phMeanDif = TRUE,
  phSig = TRUE,
  phTest = FALSE,
  phFlag = FALSE,
  formula
)
```

## Arguments

data	the data as a data frame
deps	a string naming the dependent variables in data
group	a string naming the grouping or independent variable in data
welchs	TRUE (default) or FALSE, perform Welch's one-way ANOVA which does not assume equal variances
fishers	TRUE or FALSE (default), perform Fisher's one-way ANOVA which assumes equal variances
miss	'perAnalysis' or 'listwise', how to handle missing values; 'perAnalysis' excludes missing values for individual dependent variables, 'listwise' excludes a row from all analyses if one of its entries is missing.

desc	TRUE or FALSE (default), provide descriptive statistics
descPlot	TRUE or FALSE (default), provide descriptive plots
norm	TRUE or FALSE (default), perform Shapiro-Wilk test of normality
qq	TRUE or FALSE (default), provide a Q-Q plot of residuals
eqv	TRUE or FALSE (default), perform Levene's test for homogeneity of variances
phMethod	'none', 'gamesHowell' or 'tukey', which post-hoc tests to provide; 'none' shows no post-hoc tests, 'gamesHowell' shows Games-Howell post-hoc tests where no equivalence of variances is assumed, and 'tukey' shows Tukey post-hoc tests where equivalence of variances is assumed
phMeanDiff	TRUE (default) or FALSE, provide mean differences for post-hoc tests
phSig	TRUE (default) or FALSE, provide significance levels for post-hoc tests
phTest	TRUE or FALSE (default), provide test results (t-value and degrees of freedom) for post-hoc tests
phFlag	TRUE or FALSE (default), flag significant post-hoc comparisons
formula	(optional) the formula to use, see the examples

## Details

For convenience, this method allows specifying multiple dependent variables, resulting in multiple independent tests.

Note that the Welch's ANOVA is the same procedure as the Welch's independent samples t-test.

## Value

A results object containing:

results\$anova	a table of the test results
results\$desc	a table containing the group descriptives
results\$assump\$norm	a table containing the normality tests
results\$assump\$eqv	a table of homogeneity of variances tests
results\$plots	an array of groups of plots
results\$postHoc	an array of post-hoc tables

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$anova$asDF
as.data.frame(results$anova)
```

## Examples

```
data('ToothGrowth')
dat <- ToothGrowth
dat$dose <- factor(dat$dose)

anovaOneW(formula = len ~ dose, data = dat)
```

```

#
# ONE-WAY ANOVA
#
# One-Way ANOVA (Welch's)
# -----
#      F      df1      df2      p
# -----
#   len   68.4      2    37.7 < .001
# -----
#

```

## anovaRM

*Repeated Measures ANOVA***Description**

The Repeated Measures ANOVA is used to explore the relationship between a continuous dependent variable and one or more categorical explanatory variables, where one or more of the explanatory variables are 'within subjects' (where multiple measurements are from the same subject). Additionally, this analysis allows the inclusion of covariates, allowing for repeated measures ANCOVAs as well.

**Usage**

```
anovaRM(
  data,
  rm = list(list(label = "RM Factor 1", levels = list("Level 1", "Level 2"))),
  rmCells = NULL,
  bs = NULL,
  cov = NULL,
  effectSize = NULL,
  depLabel = "Dependent",
  rmTerms = NULL,
  bsTerms = NULL,
  ss = "3",
  spherTests = FALSE,
  spherCorr = list("none"),
  leveneTest = FALSE,
  contrasts = NULL,
  postHoc = NULL,
  postHocCorr = list("tukey"),
  emMeans = list(list()),
  emmPlots = TRUE,
  emmTables = FALSE,
  emmWeights = TRUE,
  ciWidthEmm = 95,
```

```

    emmPlotData = FALSE,
    emmPlotError = "ci",
    groupSumm = FALSE
)

```

## Arguments

<code>data</code>	the data as a data frame
<code>rm</code>	a list of lists, where each list describes the <code>label</code> (as a string) and the <code>levels</code> (as vector of strings) of a particular repeated measures factor
<code>rmCells</code>	a list of lists, where each list describes a repeated measure (as a string) from <code>data</code> defined as <code>measure</code> and the particular combination of levels from <code>rm</code> that it belongs to (as a vector of strings) defined as <code>cell</code>
<code>bs</code>	a vector of strings naming the between subjects factors from <code>data</code>
<code>cov</code>	a vector of strings naming the covariates from <code>data</code> . Variables must be numeric
<code>effectSize</code>	one or more of 'eta', 'partEta', or 'omega'; use eta <sup>2</sup> , partial eta <sup>2</sup> , and omega <sup>2</sup> effect sizes, respectively
<code>depLabel</code>	a string (default: 'Dependent') describing the label used for the dependent variable throughout the analysis
<code>rmTerms</code>	a list of character vectors describing the repeated measures terms to go into the model
<code>bsTerms</code>	a list of character vectors describing the between subjects terms to go into the model
<code>ss</code>	'2' or '3' (default), the sum of squares to use
<code>spherTests</code>	TRUE or FALSE (default), perform sphericity tests
<code>spherCorr</code>	one or more of 'none' (default), 'GG', or HF; use no p-value correction, the Greenhouse-Geisser p-value correction, and the Huynh-Feldt p-value correction for shericity, respectively
<code>leveneTest</code>	TRUE or FALSE (default), test for homogeneity of variances (i.e., Levene's test)
<code>contrasts</code>	in development
<code>postHoc</code>	a list of character vectors describing the post-hoc tests that need to be computed
<code>postHocCorr</code>	one or more of 'none', 'tukey' (default), 'scheffe', 'bonf', or 'holm'; use no, Tukey, Scheffe, Bonferroni and Holm posthoc corrections, respectively
<code>emMeans</code>	a list of lists specifying the variables for which the estimated marginal means need to be calculate. Supports up to three variables per term.
<code>emmPlots</code>	TRUE (default) or FALSE, provide estimated marginal means plots
<code>emmTables</code>	TRUE or FALSE (default), provide estimated marginal means tables
<code>emmWeights</code>	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency
<code>ciWidthEmm</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
<code>emmPlotData</code>	TRUE or FALSE (default), plot the data on top of the marginal means
<code>emmPlotError</code>	'none', 'ci' (default), or 'se'. Use no error bars, use confidence intervals, or use standard errors on the marginal mean plots, respectively
<code>groupSumm</code>	TRUE or FALSE (default), report a summary of the different groups

## Details

This analysis requires that the data be in 'wide format', where each row represents a subject (as opposed to long format, where each measurement of the dependent variable is represented as a row).

A non-parametric equivalent of the repeated measures ANOVA also exists; the Friedman test. However, it has the limitation of only being able to test a single factor.

## Value

A results object containing:

results\$rmTable	a table
results\$bsTable	a table
results\$assump\$spherTable	a table
results\$assump\$leveneTable	a table
results\$contrasts	an array of tables
results\$postHoc	an array of tables
results\$emm	an array of the estimated marginal means plots + tables
results\$groupSummary	a summary of the groups

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$rmTable$asDF
as.data.frame(results$rmTable)
```

## Examples

## Not run:

```
data('bugs', package = 'jmv')

anovaRM(
  data = bugs,
  rm = list(
    list(
      label = 'Frightening',
      levels = c('Low', 'High'))),
  rmCells = list(
    list(
      measure = 'LDLF',
      cell = 'Low'),
    list(
      measure = 'LDHF',
      cell = 'High')),
  rmTerms = list(
    'Frightening'))

#
# REPEATED MEASURES ANOVA
```

```

#
# Within Subjects Effects
# -----
#          Sum of Squares   df   Mean Square   F    p
# -----
#   Frightening        126    1      126.11  44.2 < .001
#   Residual          257   90       2.85
# -----
#   Note. Type 3 Sums of Squares
#
#
# Between Subjects Effects
# -----
#          Sum of Squares   df   Mean Square   F    p
# -----
#   Residual          954   90       10.6
# -----
#   Note. Type 3 Sums of Squares
#
## End(Not run)

```

## Description

The Friedman test is used to explore the relationship between a continuous dependent variable and a categorical explanatory variable, where the explanatory variable is 'within subjects' (where multiple measurements are from the same subject). It is analogous to Repeated Measures ANOVA, but with the advantage of being non-parametric, and not requiring the assumptions of normality or homogeneity of variances. However, it has the limitation that it can only test a single explanatory variable at a time.

## Usage

```

anovaRMNP(
  data,
  measures,
  pairs = FALSE,
  desc = FALSE,
  plots = FALSE,
  plotType = "means"
)

```

### Arguments

data	the data as a data frame
measures	a vector of strings naming the repeated measures variables
pairs	TRUE or FALSE (default), perform pairwise comparisons
desc	TRUE or FALSE (default), provide descriptive statistics
plots	TRUE or FALSE (default), provide a descriptive plot
plotType	'means' (default) or 'medians', the error bars to use in the plot

### Value

A results object containing:

results\$table	a table of the Friedman test results
results\$comp	a table of the pairwise comparisons
results\$desc	a table containing the descriptives
results\$plot	a descriptives plot

Tables can be converted to data frames with asDF or [as.data.frame](#). For example:

```
results$table$asDF
as.data.frame(results$table)
```

### Examples

```
data('bugs', package = 'jmv')

anovaRMNP(bugs, measures = vars(LDLF, LDHF, HDLF, HDHF))

#
# REPEATED MEASURES ANOVA (NON-PARAMETRIC)
#
# Friedman
# -----
#   X2      df      p
# -----
#   55.8      3    < .001
# -----
```

### Description

data sets

**Author(s)**

Ryan, Wilde & Crist (2013)

**References**

<http://faculty.kutztown.edu/rryan/RESEARCH/PUBS/Ryan,%20Wilde,%20%26%20Crist%202013%20Web%20exp%20vs%20lab.pdf>

---

cfa

*Confirmatory Factor Analysis*

---

**Description**

Confirmatory Factor Analysis

**Usage**

```
cfa(
  data,
  factors = list(list(label = "Factor 1", vars = list())),
  resCov,
  miss = "fiml",
  constrain = "facVar",
  estTest = TRUE,
  ci = FALSE,
  ciWidth = 95,
  stdEst = FALSE,
  factCovEst = TRUE,
  factInterceptEst = FALSE,
  resCovEst = FALSE,
  resInterceptEst = FALSE,
  fitMeasures = list("cfi", "tli", "rmsea"),
  modelTest = TRUE,
  pathDiagram = FALSE,
  corRes = FALSE,
  hlCorRes = 0.1,
  mi = FALSE,
  hlMI = 3
)
```

**Arguments**

data	the data as a data frame
factors	a list containing named lists that define the <code>label</code> of the factor and the <code>vars</code> that belong to that factor
resCov	a list of lists specifying the residual covariances that need to be estimated

miss	'listwise' or 'fiml', how to handle missing values; 'listwise' excludes a row from all analyses if one of its entries is missing, 'fiml' uses a full information maximum likelihood method to estimate the model.
constraint	'facVar' or 'facInd', how to constrain the model; 'facVar' fixes the factor variances to one, 'facInd' fixes each factor to the scale of its first indicator.
estTest	TRUE (default) or FALSE, provide 'Z' and 'p' values for the model estimates
ci	TRUE or FALSE (default), provide a confidence interval for the model estimates
ciWidth	a number between 50 and 99.9 (default: 95) specifying the confidence interval width that is used as 'ci'
stdEst	TRUE or FALSE (default), provide a standardized estimate for the model estimates
factCovEst	TRUE (default) or FALSE, provide estimates for the factor (co)variances
factInterceptEst	TRUE or FALSE (default), provide estimates for the factor intercepts
resCovEst	TRUE (default) or FALSE, provide estimates for the residual (co)variances
resInterceptEst	TRUE or FALSE (default), provide estimates for the residual intercepts
fitMeasures	one or more of 'cfi', 'tli', 'srmr', 'rmsea', 'aic', or 'bic'; use CFI, TLI, SRMR, RMSEA + 90% confidence interval, adjusted AIC, and BIC model fit measures, respectively
modelTest	TRUE (default) or FALSE, provide a chi-square test for exact fit that compares the model with the perfect fitting model
pathDiagram	TRUE or FALSE (default), provide a path diagram of the model
corRes	TRUE or FALSE (default), provide the residuals for the observed correlation matrix (i.e., the difference between the expected correlation matrix and the observed correlation matrix)
hlCorRes	a number (default: 0.1), highlight values in the 'corRes' table above this value
mi	TRUE or FALSE (default), provide modification indices for the parameters not included in the model
hlMI	a number (default: 3), highlight values in the 'modIndices' tables above this value

## Value

A results object containing:

results\$factorLoadings	a table containing the factor loadings
results\$factorEst\$factorCov	a table containing factor covariances estimates
results\$factorEst\$factorIntercept	a table containing factor intercept estimates
results\$resEst\$resCov	a table containing residual covariances estimates
results\$resEst\$resIntercept	a table containing residual intercept estimates
results\$modelFit\$test	a table containing the chi-square test for exact fit
results\$modelFit\$fitMeasures	a table containing fit measures
results\$modelPerformance\$corRes	a table containing residuals for the observed correlation matrix
results\$modelPerformance\$modIndices	a group
results\$pathDiagram	an image containing the model path diagram
results\$modelSyntax	the lavaan syntax used to fit the model

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$factorLoadings$asDF
as.data.frame(results$factorLoadings)
```

## Examples

```
data <- lavaan::HolzingerSwineford1939

jmv:::cfa(
  data = data,
  factors = list(
    list(label="Visual", vars=c("x1", "x2", "x3")),
    list(label="Textual", vars=c("x4", "x5", "x6")),
    list(label="Speed", vars=c("x7", "x8", "x9"))),
  resCov = NULL)

#
# CONFIRMATORY FACTOR ANALYSIS
#
# Factor Loadings
# -----
#   Factor   Indicator   Estimate     SE      Z      p
# -----
#   Visual     x1        0.900  0.0832  10.81  < .001
#             x2        0.498  0.0808   6.16  < .001
#             x3        0.656  0.0776   8.46  < .001
#   Textual    x4        0.990  0.0567  17.46  < .001
#             x5        1.102  0.0626  17.60  < .001
#             x6        0.917  0.0538  17.05  < .001
#   Speed      x7        0.619  0.0743   8.34  < .001
#             x8        0.731  0.0755   9.68  < .001
#             x9        0.670  0.0775   8.64  < .001
# -----
#
#
# FACTOR ESTIMATES
#
# Factor Covariances
# -----
#           Estimate     SE      Z      p
# -----
#   Visual   Visual  1.000 a
#             Textual  0.459  0.0635   7.22  < .001
#             Speed   0.471  0.0862   5.46  < .001
#   Textual  Textual 1.000 a
#             Speed   0.283  0.0715   3.96  < .001
#   Speed    Speed  1.000 a
# -----
#
#   a fixed parameter
#
#
# MODEL FIT
```

```
#  
# Test for Exact Fit  
# -----  
# X2      df      p  
# -----  
# 85.3     24    < .001  
# -----  
#  
#  
# Fit Measures  
# -----  
# CFI      TLI      RMSEA    Lower    Upper  
# -----  
# 0.931    0.896    0.0921   0.0714   0.114  
# -----  
#
```

---

**contTables***Contingency Tables*

---

**Description**

The  $X^2$  test of association (not to be confused with the  $X^2$  goodness of fit) is used to test whether two categorical variables are independent or associated. If the p-value is low, it suggests the variables are not independent, and that there is a relationship between the two variables.

**Usage**

```
contTables(  
  data,  
  rows,  
  cols,  
  counts = NULL,  
  layers = NULL,  
  chiSq = TRUE,  
  chiSqCorr = FALSE,  
  likeRat = FALSE,  
  fisher = FALSE,  
  contCoef = FALSE,  
  phiCra = FALSE,  
  logOdds = FALSE,  
  odds = FALSE,  
  relRisk = FALSE,  
  ci = TRUE,  
  ciWidth = 95,  
  gamma = FALSE,  
  taub = FALSE,
```

```

obs = TRUE,
exp = FALSE,
pcRow = FALSE,
pcCol = FALSE,
pcTot = FALSE,
formula
)

```

### Arguments

<code>data</code>	the data as a data frame
<code>rows</code>	the variable to use as the rows in the contingency table (not necessary when providing a formula, see the examples)
<code>cols</code>	the variable to use as the columns in the contingency table (not necessary when providing a formula, see the examples)
<code>counts</code>	the variable to use as the counts in the contingency table (not necessary when providing a formula, see the examples)
<code>layers</code>	the variables to use to split the contingency table (not necessary when providing a formula, see the examples)
<code>chiSq</code>	TRUE (default) or FALSE, provide $\chi^2$
<code>chiSqCorr</code>	TRUE or FALSE (default), provide $\chi^2$ with continuity correction
<code>likeRat</code>	TRUE or FALSE (default), provide the likelihood ratio
<code>fisher</code>	TRUE or FALSE (default), provide Fisher's exact test
<code>contCoef</code>	TRUE or FALSE (default), provide the contingency coefficient
<code>phiCra</code>	TRUE or FALSE (default), provide Phi and Cramer's V
<code>logOdds</code>	TRUE or FALSE (default), provide the log odds ratio (only available for 2x2 tables)
<code>odds</code>	TRUE or FALSE (default), provide the odds ratio (only available for 2x2 tables)
<code>relRisk</code>	TRUE or FALSE (default), provide the relative risk (only available for 2x2 tables)
<code>ci</code>	TRUE or FALSE (default), provide confidence intervals for the comparative measures
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95), width of the confidence intervals to provide
<code>gamma</code>	TRUE or FALSE (default), provide gamma
<code>taub</code>	TRUE or FALSE (default), provide Kendall's tau-b
<code>obs</code>	TRUE or FALSE (default), provide the observed counts
<code>exp</code>	TRUE or FALSE (default), provide the expected counts
<code>pcRow</code>	TRUE or FALSE (default), provide row percentages
<code>pcCol</code>	TRUE or FALSE (default), provide column percentages
<code>pcTot</code>	TRUE or FALSE (default), provide total percentages
<code>formula</code>	(optional) the formula to use, see the examples

### Value

A results object containing:

results\$freqs	a table of proportions
results\$chiSq	a table of $\chi^2$ test results
results\$odds	a table of comparative measures
results\$nom	a table of the 'nominal' test results
results\$gamma	a table of the gamma test results
results\$taub	a table of the Kendall's tau-b test results

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$freqs$asDF
as.data.frame(results$freqs)
```

## Examples

```
data('HairEyeColor')
dat <- as.data.frame(HairEyeColor)

contTables(formula = Freq ~ Hair:Eye, dat)

#
#  CONTINGENCY TABLES
#
#  Contingency Tables
#  -----
#    Hair   Brown   Blue   Hazel   Green   Total
#  -----
#    Black      68      20      15       5     108
#    Brown     119      84      54      29     286
#    Red        26      17      14      14      71
#    Blond       7      94      10      16     127
#    Total     220     215     93      64     592
#  -----
#
#
#  X2 Tests
#  -----
#          Value   df     p
#  -----
#    X2      138     9   < .001
#    N        592
#  -----
#
# Alternatively, omit the left of the formula ('Freq') if each row
# represents a single observation:

contTables(formula = ~ Hair:Eye, dat)
```

---

contTablesPaired*Paired Samples Contingency Tables*

---

## Description

McNemar test

## Usage

```
contTablesPaired(
  data,
  rows,
  cols,
  counts = NULL,
  chiSq = TRUE,
  chiSqCorr = FALSE,
  exact = FALSE,
  pcRow = FALSE,
  pcCol = FALSE,
  formula
)
```

## Arguments

data	the data as a data frame
rows	the variable to use as the rows in the contingency table (not necessary when providing a formula, see the examples)
cols	the variable to use as the columns in the contingency table (not necessary when providing a formula, see the examples)
counts	the variable to use as the counts in the contingency table (not necessary when providing a formula, see the examples)
chiSq	TRUE (default) or FALSE, provide $\chi^2$
chiSqCorr	TRUE or FALSE (default), provide $\chi^2$ with continuity correction
exact	TRUE or FALSE (default), provide an exact log odds ratio (requires exact2x2 to be installed)
pcRow	TRUE or FALSE (default), provide row percentages
pcCol	TRUE or FALSE (default), provide column percentages
formula	(optional) the formula to use, see the examples

## Value

A results object containing:

results\$freqs	a proportions table
results\$test	a table of test results

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$freqs$asDF
as.data.frame(results$freqs)
```

## Examples

```
dat <- data.frame(
  `1st survey` = c('Approve', 'Approve', 'Disapprove', 'Disapprove'),
  `2nd survey` = c('Approve', 'Disapprove', 'Approve', 'Disapprove'),
  `Counts` = c(794, 150, 86, 570),
  check.names=FALSE)

contTablesPaired(formula = Counts ~ `1st survey`:`2nd survey`, data = dat)

#
# PAIRED SAMPLES CONTINGENCY TABLES
#
# Contingency Tables
# -----
#   1st survey    Approve   Disapprove   Total
# -----
#   Approve        794        150        944
#   Disapprove      86        570        656
#   Total          880        720       1600
# -----
#
#
# McNemar Test
# -----
#                   Value     df      p
# -----
#   X2            17.4      1    < .001
#   X2 continuity correction  16.8      1    < .001
# -----
```

---

# Alternatively, omit the left of the formula (`Counts`) from the  
# formula if each row represents a single observation:

```
contTablesPaired(formula = ~ `1st survey`:`2nd survey`, data = dat)
```

---

## Description

Correlation matrices are a way to examine linear relationships between two or more continuous variables.

**Usage**

```
corrMatrix(
  data,
  vars,
  pearson = TRUE,
  spearman = FALSE,
  kendall = FALSE,
  sig = TRUE,
  flag = FALSE,
  n = FALSE,
  ci = FALSE,
  ciWidth = 95,
  plots = FALSE,
  plotDens = FALSE,
  plotStats = FALSE,
  hypothesis = "corr"
)
```

**Arguments**

<code>data</code>	the data as a data frame
<code>vars</code>	a vector of strings naming the variables to correlate in <code>data</code>
<code>pearson</code>	TRUE (default) or FALSE, provide Pearson's R
<code>spearman</code>	TRUE or FALSE (default), provide Spearman's rho
<code>kendall</code>	TRUE or FALSE (default), provide Kendall's tau-b
<code>sig</code>	TRUE (default) or FALSE, provide significance levels
<code>flag</code>	TRUE or FALSE (default), flag significant correlations
<code>n</code>	TRUE or FALSE (default), provide the number of cases
<code>ci</code>	TRUE or FALSE (default), provide confidence intervals
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95), the width of confidence intervals to provide
<code>plots</code>	TRUE or FALSE (default), provide a correlation matrix plot
<code>plotDens</code>	TRUE or FALSE (default), provide densities in the correlation matrix plot
<code>plotStats</code>	TRUE or FALSE (default), provide statistics in the correlation matrix plot
<code>hypothesis</code>	one of 'corr' (default), 'pos', 'neg' specifying the alternative hypothesis; correlated, correlated positively, correlated negatively respectively.

**Details**

For each pair of variables, a Pearson's r value indicates the strength and direction of the relationship between those two variables. A positive value indicates a positive relationship (higher values of one variable predict higher values of the other variable). A negative Pearson's r indicates a negative relationship (higher values of one variable predict lower values of the other variable, and vice-versa). A value of zero indicates no relationship (whether a variable is high or low, does not tell us anything about the value of the other variable).

More formally, it is possible to test the null hypothesis that the correlation is zero and calculate a p-value. If the p-value is low, it suggests the correlation co-efficient is not zero, and there is a linear (or more complex) relationship between the two variables.

### Value

A results object containing:

results\$matrix	a correlation matrix table
results\$plot	a correlation matrix plot

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$matrix$asDF
as.data.frame(results$matrix)
```

### Examples

```
## Not run:
data('mtcars')

corrMatrix(mtcars, vars = vars(mpg, cyl, disp, hp))

#
# CORRELATION MATRIX
#
# Correlation Matrix
# -----
#          mpg     cyl      disp      hp
# -----
#   mpg    Pearson's r      -0.852   -0.848   -0.776
#          p-value           < .001    < .001    < .001
#
#   cyl    Pearson's r       0.902    0.832
#          p-value           < .001    < .001
#
#   disp   Pearson's r       0.791
#          p-value           < .001
#
#   hp     Pearson's r
#          p-value
# -----
#
## End(Not run)
```

---

descriptives	<i>Descriptives</i>
--------------	---------------------

---

## Description

Descriptives are an assortment of summarising statistics, and visualizations which allow exploring the shape and distribution of data. It is good practice to explore your data with descriptives before proceeding to more formal tests.

## Usage

```
descriptives(  
  data,  
  vars,  
  splitBy = NULL,  
  freq = FALSE,  
  hist = FALSE,  
  dens = FALSE,  
  bar = FALSE,  
  barCounts = FALSE,  
  box = FALSE,  
  violin = FALSE,  
  dot = FALSE,  
  dotType = "jitter",  
  qq = FALSE,  
  n = TRUE,  
  missing = TRUE,  
  mean = TRUE,  
  median = TRUE,  
  mode = FALSE,  
  sum = FALSE,  
  sd = FALSE,  
  variance = FALSE,  
  range = FALSE,  
  min = TRUE,  
  max = TRUE,  
  se = FALSE,  
  skew = FALSE,  
  kurt = FALSE,  
  sw = FALSE,  
  quart = FALSE,  
  pcEqGr = FALSE,  
  pcNEqGr = 4,  
  formula  
)
```

## Arguments

data	the data as a data frame
vars	a vector of strings naming the variables of interest in data
splitBy	a vector of strings naming the variables used to split vars
freq	TRUE or FALSE (default), provide frequency tables (nominal, ordinal variables only)
hist	TRUE or FALSE (default), provide histograms (continuous variables only)
dens	TRUE or FALSE (default), provide density plots (continuous variables only)
bar	TRUE or FALSE (default), provide bar plots (nominal, ordinal variables only)
barCounts	TRUE or FALSE (default), add counts to the bar plots
box	TRUE or FALSE (default), provide box plots (continuous variables only)
violin	TRUE or FALSE (default), provide violin plots (continuous variables only)
dot	TRUE or FALSE (default), provide dot plots (continuous variables only)
dotType	.
qq	TRUE or FALSE (default), provide Q-Q plots (continuous variables only)
n	TRUE (default) or FALSE, provide the sample size
missing	TRUE (default) or FALSE, provide the number of missing values
mean	TRUE (default) or FALSE, provide the mean
median	TRUE (default) or FALSE, provide the median
mode	TRUE or FALSE (default), provide the mode
sum	TRUE or FALSE (default), provide the sum
sd	TRUE or FALSE (default), provide the standard deviation
variance	TRUE or FALSE (default), provide the variance
range	TRUE or FALSE (default), provide the range
min	TRUE or FALSE (default), provide the minimum
max	TRUE or FALSE (default), provide the maximum
se	TRUE or FALSE (default), provide the standard error
skew	TRUE or FALSE (default), provide the skewness
kurt	TRUE or FALSE (default), provide the kurtosis
sw	TRUE or FALSE (default), provide Shapiro-Wilk p-value
quart	TRUE or FALSE (default), provide quartiles
pcEqGr	TRUE or FALSE (default), provide quantiles
pcNEqGr	an integer (default: 4) specifying the number of equal groups
formula	(optional) the formula to use, see the examples

## Value

A results object containing:

<code>results\$descriptives</code>	a table of the descriptive statistics
<code>results\$frequencies</code>	an array of frequency tables
<code>results\$plots</code>	an array of descriptive plots

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$descriptives$asDF
as.data.frame(results$descriptives)
```

## Examples

```
## Not run:
data('mtcars')
dat <- mtcars

# frequency tables can be provided for factors
dat$gear <- as.factor(dat$gear)

descriptives(dat, vars = vars(mpg, cyl, disp, gear), freq = TRUE)

#
# DESCRIPTIVES
#
# Descriptives
# -----
#          mpg      cyl      disp     gear
# -----
#   N        32       32       32       32
#   Missing    0       0       0       0
#   Mean      20.1    6.19    231     3.69
#   Median     19.2    6.00    196     4.00
#   Minimum    10.4    4.00    71.1      3
#   Maximum    33.9    8.00    472      5
# -----
#
#
# FREQUENCIES
#
# Frequencies of gear
# -----
#   Levels   Counts
# -----
#     3         15
#     4         12
#     5          5
# -----
#
#
# splitting by a variable
descriptives(formula = disp ~ mpg ~ cyl, dat,
            median=F, min=F, max=F, n=F, missing=F)
```

```
# providing histograms  
descriptives(formula = mpg ~ cyl, dat, hist=T,  
    median=F, min=F, max=F, n=F, missing=F)  
  
# splitting by multiple variables  
descriptives(formula = mpg ~ cyl:gear, dat,  
    median=F, min=F, max=F, missing=F)  
  
## End(Not run)
```

---

efa

*Exploratory Factor Analysis*

---

## Description

Exploratory Factor Analysis

## Usage

```
efa(  
    data,  
    vars,  
    nFactorMethod = "parallel",  
    nFactors = 1,  
    minEigen = 0,  
    extraction = "minres",  
    rotation = "oblimin",  
    hideLoadings = 0.3,  
    sortLoadings = FALSE,  
    screePlot = FALSE,  
    eigen = FALSE,  
    factorCor = FALSE,  
    factorSummary = FALSE,  
    modelFit = FALSE,  
    kmo = FALSE,  
    bartlett = FALSE  
)
```

## Arguments

data	the data as a data frame
vars	a vector of strings naming the variables of interest in <code>data</code>
nFactorMethod	'parallel' (default), 'eigen' or 'fixed', the way to determine the number of factors
nFactors	an integer (default: 1), the number of factors in the model

minEigen	a number (default: 0), the minimal eigenvalue for a factor to be included in the model
extraction	'minres' (default), 'ml', or 'pa' use respectively 'minimum residual', 'maximum likelihood', or 'principal axis' as the factor extraction method
rotation	'none', 'varimax', 'quartimax', 'promax', 'oblimin' (default), or 'simplimax', the rotation to use in estimation
hideLoadings	a number (default: 0.3), hide factor loadings below this value
sortLoadings	TRUE or FALSE (default), sort the factor loadings by size
screePlot	TRUE or FALSE (default), show scree plot
eigen	TRUE or FALSE (default), show eigenvalue table
factorCor	TRUE or FALSE (default), show factor correlations
factorSummary	TRUE or FALSE (default), show factor summary
modelFit	TRUE or FALSE (default), show model fit measures and test
kmo	TRUE or FALSE (default), show Kaiser-Meyer-Olkin (KMO) measure of sampling adequacy (MSA) results
bartlett	TRUE or FALSE (default), show Bartlett's test of sphericity results

### Value

A results object containing:

results\$text	a preformatted
---------------	----------------

### Examples

```

data('iris')

efa(iris, vars = vars(Sepal.Length, Sepal.Width, Petal.Length, Petal.Width))

#
# EXPLORATORY FACTOR ANALYSIS
#
# Factor Loadings
# -----
#          1       2      Uniqueness
# -----
# Sepal.Length  0.993    0.10181
# Sepal.Width   0.725    0.42199
# Petal.Length  0.933    0.00483
# Petal.Width   0.897    0.07088
# -----
# Note. 'oblimin' rotation was used
#

```

---

linReg*Linear Regression*

---

**Description**

Linear regression is used to explore the relationship between a continuous dependent variable, and one or more continuous and/or categorical explanatory variables. Other statistical methods, such as ANOVA and ANCOVA, are in reality just forms of linear regression.

**Usage**

```
linReg(  
  data,  
  dep,  
  covs = NULL,  
  factors = NULL,  
  blocks = list(list()),  
  refLevels = NULL,  
  intercept = "refLevel",  
  r = TRUE,  
  r2 = TRUE,  
  r2Adj = FALSE,  
  aic = FALSE,  
  bic = FALSE,  
  rmse = FALSE,  
  modelTest = FALSE,  
  anova = FALSE,  
  ci = FALSE,  
  ciWidth = 95,  
  stdEst = FALSE,  
  ciStdEst = FALSE,  
  ciWidthStdEst = 95,  
  coefPlot = FALSE,  
  norm = FALSE,  
  qqPlot = FALSE,  
  resPlots = FALSE,  
  durbin = FALSE,  
  collin = FALSE,  
  cooks = FALSE,  
  emMeans = list(list()),  
  ciEmm = TRUE,  
  ciWidthEmm = 95,  
  emmPlots = TRUE,  
  emmTables = FALSE,  
  emmWeights = TRUE  
)
```

### Arguments

<code>data</code>	the data as a data frame
<code>dep</code>	the dependent variable from <code>data</code> , variable must be numeric
<code>covs</code>	the covariates from <code>data</code>
<code>factors</code>	the fixed factors from <code>data</code>
<code>blocks</code>	a list containing vectors of strings that name the predictors that are added to the model. The elements are added to the model according to their order in the list
<code>refLevels</code>	a list of lists specifying reference levels of the dependent variable and all the factors
<code>intercept</code>	'refLevel' (default) or 'grandMean', coding of the intercept. Either creates contrast so that the intercept represents the reference level or the grand mean
<code>r</code>	TRUE (default) or FALSE, provide the statistical measure R for the models
<code>r2</code>	TRUE (default) or FALSE, provide the statistical measure R-squared for the models
<code>r2Adj</code>	TRUE or FALSE (default), provide the statistical measure adjusted R-squared for the models
<code>aic</code>	TRUE or FALSE (default), provide Aikaike's Information Criterion (AIC) for the models
<code>bic</code>	TRUE or FALSE (default), provide Bayesian Information Criterion (BIC) for the models
<code>rmse</code>	TRUE or FALSE (default), provide RMSE for the models
<code>modelTest</code>	TRUE (default) or FALSE, provide the model comparison between the models and the NULL model
<code>anova</code>	TRUE or FALSE (default), provide the omnibus ANOVA test for the predictors
<code>ci</code>	TRUE or FALSE (default), provide a confidence interval for the model coefficients
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
<code>stdEst</code>	TRUE or FALSE (default), provide a standardized estimate for the model coefficients
<code>ciStdEst</code>	TRUE or FALSE (default), provide a confidence interval for the model coefficient standardized estimates
<code>ciWidthStdEst</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
<code>coefPlot</code>	TRUE or FALSE (default), provide a coefficient plot where for each predictor the estimated coefficient and confidence intervals are plotted.
<code>norm</code>	TRUE or FALSE (default), perform a Shapiro-Wilk test on the residuals
<code>qqPlot</code>	TRUE or FALSE (default), provide a Q-Q plot of residuals
<code>resPlots</code>	TRUE or FALSE (default), provide residual plots where the dependent variable and each covariate is plotted against the standardized residuals.
<code>durbin</code>	TRUE or FALSE (default), provide results of the Durbin- Watson test for autocorrelation

collin	TRUE or FALSE (default), provide VIF and tolerance collinearity statistics
cooks	TRUE or FALSE (default), provide summary statistics for the Cook's distance
emMeans	a formula containing the terms to estimate marginal means for, supports up to three variables per term
ciEmm	TRUE (default) or FALSE, provide a confidence interval for the estimated marginal means
ciWidthEmm	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
emmPlots	TRUE (default) or FALSE, provide estimated marginal means plots
emmTables	TRUE or FALSE (default), provide estimated marginal means tables
emmWeights	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency

### Value

A results object containing:

results\$modelFit	a table
results\$modelComp	a table
results\$models	an array of model specific results

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$modelFit$asDF
as.data.frame(results$modelFit)
```

### Examples

```
data('Prestige', package='carData')

linReg(data = Prestige, dep = income,
       covs = vars(education, prestige, women),
       blocks = list(list('education', 'prestige', 'women')))

#
# LINEAR REGRESSION
#
# Model Fit Measures
# -----
#   Model     R      R^2
# -----
#   1    0.802    0.643
# -----
#
#
# MODEL SPECIFIC RESULTS
#
```

```

# MODEL 1
#
#
# Model Coefficients
# -----
#   Predictor   Estimate    SE      t      p
# -----
#   Intercept   -253.8   1086.16  -0.234   0.816
#   women       -50.9     8.56   -5.948   < .001
#   prestige     141.4    29.91    4.729   < .001
#   education   177.2    187.63    0.944   0.347
# -----
#

```

**logLinear***Log-Linear Regression***Description**

Log-Linear Regression

**Usage**

```

logLinear(
  data,
  factors = NULL,
  counts = NULL,
  blocks = list(list()),
  refLevels = NULL,
  modelTest = FALSE,
  dev = TRUE,
  aic = TRUE,
  bic = FALSE,
  pseudoR2 = list("r2mf"),
  omni = FALSE,
  ci = FALSE,
  ciWidth = 95,
  RR = FALSE,
  ciRR = FALSE,
  ciWidthRR = 95,
  emMeans = list(list()),
  ciEmm = TRUE,
  ciWidthEmm = 95,
  emmPlots = TRUE,
  emmTables = FALSE,
  emmWeights = TRUE
)

```

**Arguments**

<b>data</b>	the data as a data frame
<b>factors</b>	a vector of strings naming the factors from <b>data</b>
<b>counts</b>	a string naming a variable in <b>data</b> containing counts, or NULL if each row represents a single observation
<b>blocks</b>	a list containing vectors of strings that name the predictors that are added to the model. The elements are added to the model according to their order in the list
<b>refLevels</b>	a list of lists specifying reference levels of the dependent variable and all the factors
<b>modelTest</b>	TRUE or FALSE (default), provide the model comparison between the models and the NULL model
<b>dev</b>	TRUE (default) or FALSE, provide the deviance (or -2LogLikelihood) for the models
<b>aic</b>	TRUE (default) or FALSE, provide Aikaike's Information Criterion (AIC) for the models
<b>bic</b>	TRUE or FALSE (default), provide Bayesian Information Criterion (BIC) for the models
<b>pseudoR2</b>	one or more of 'r2mf', 'r2cs', or 'r2n'; use McFadden's, Cox & Snell, and Nagelkerke pseudo-R <sup>2</sup> , respectively
<b>omni</b>	TRUE or FALSE (default), provide the omnibus likelihood ratio tests for the predictors
<b>ci</b>	TRUE or FALSE (default), provide a confidence interval for the model coefficient estimates
<b>ciWidth</b>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
<b>RR</b>	TRUE or FALSE (default), provide the exponential of the log-rate ratio estimate, or the rate ratio estimate
<b>ciRR</b>	TRUE or FALSE (default), provide a confidence interval for the model coefficient rate ratio estimates
<b>ciWidthRR</b>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
<b>emMeans</b>	a list of lists specifying the variables for which the estimated marginal means need to be calculate. Supports up to three variables per term.
<b>ciEmm</b>	TRUE (default) or FALSE, provide a confidence interval for the estimated marginal means
<b>ciWidthEmm</b>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
<b>emmPlots</b>	TRUE (default) or FALSE, provide estimated marginal means plots
<b>emmTables</b>	TRUE or FALSE (default), provide estimated marginal means tables
<b>emmWeights</b>	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency

**Value**

A results object containing:

results\$modelFit	a table
results\$modelComp	a table
results\$models	an array of model specific results

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$modelFit$asDF
as.data.frame(results$modelFit)
```

## Examples

```
data('mtcars')

tab <- table('gear'=mtcars$gear, 'cyl'=mtcars$cyl)
dat <- as.data.frame(tab)

logLinear(data = dat, factors = vars(gear, cyl), counts = Freq,
          blocks = list(list("gear", "cyl", c("gear", "cyl"))),
          refLevels = list(
            list(var="gear", ref="3"),
            list(var="cyl", ref="4"))

#
# LOG-LINEAR REGRESSION
#
# Model Fit Measures
# -----
#   Model    Deviance    AIC    R^2-McF
# -----
#     1    4.12e-10    41.4    1.000
# -----
#
#
# MODEL SPECIFIC RESULTS
#
# MODEL 1
#
# Model Coefficients
# -----
#   Predictor      Estimate      SE       Z       p
# -----
#   Intercept    -4.71e-16    1.00  -4.71e-16  1.000
#   gear:
#     4 3           2.079      1.06    1.961    0.050
#     5 3           0.693      1.22    0.566    0.571
#   cyl:
#     6 4           0.693      1.22    0.566    0.571
#     8 4           2.485      1.04    2.387    0.017
#   gear:cyl:
#     (4 3):(6 4)  -1.386      1.37   -1.012    0.311
#     (5 3):(6 4)  -1.386      1.73   -0.800    0.423
```

```
#   (4 3):(8 4)      -26.867    42247.17    -6.36e -4    0.999
#   (5 3):(8 4)      -2.485       1.44       -1.722    0.085
# -----
#
```

**logRegBin***Binomial Logistic Regression***Description**

Binomial Logistic Regression

**Usage**

```
logRegBin(
  data,
  dep,
  covs = NULL,
  factors = NULL,
  blocks = list(list()),
  refLevels = NULL,
  modelTest = FALSE,
  dev = TRUE,
  aic = TRUE,
  bic = FALSE,
  pseudoR2 = list("r2mf"),
  omni = FALSE,
  ci = FALSE,
  ciWidth = 95,
  OR = FALSE,
  ciOR = FALSE,
  ciWidthOR = 95,
  emMeans = list(list()),
  ciEmm = TRUE,
  ciWidthEmm = 95,
  emmPlots = TRUE,
  emmTables = FALSE,
  emmWeights = TRUE,
  class = FALSE,
  acc = FALSE,
  spec = FALSE,
  sens = FALSE,
  auc = FALSE,
  rocPlot = FALSE,
  cutOff = 0.5,
```

```

    cutOffPlot = FALSE,
    collin = FALSE,
    boxTidwell = FALSE,
    cooks = FALSE
)

```

## Arguments

data	the data as a data frame
dep	a string naming the dependent variable from data, variable must be a factor
covs	a vector of strings naming the covariates from data
factors	a vector of strings naming the fixed factors from data
blocks	a list containing vectors of strings that name the predictors that are added to the model. The elements are added to the model according to their order in the list
refLevels	a list of lists specifying reference levels of the dependent variable and all the factors
modelTest	TRUE or FALSE (default), provide the model comparison between the models and the NULL model
dev	TRUE (default) or FALSE, provide the deviance (or -2LogLikelihood) for the models
aic	TRUE (default) or FALSE, provide Aikaike's Information Criterion (AIC) for the models
bic	TRUE or FALSE (default), provide Bayesian Information Criterion (BIC) for the models
pseudoR2	one or more of 'r2mf', 'r2cs', or 'r2n'; use McFadden's, Cox & Snell, and Nagelkerke pseudo-R <sup>2</sup> , respectively
omni	TRUE or FALSE (default), provide the omnibus likelihood ratio tests for the predictors
ci	TRUE or FALSE (default), provide a confidence interval for the model coefficient estimates
ciWidth	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
OR	TRUE or FALSE (default), provide the exponential of the log-odds ratio estimate, or the odds ratio estimate
ciOR	TRUE or FALSE (default), provide a confidence interval for the model coefficient odds ratio estimates
ciWidthOR	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
emMeans	a list of lists specifying the variables for which the estimated marginal means need to be calculate. Supports up to three variables per term.
ciEmm	TRUE (default) or FALSE, provide a confidence interval for the estimated marginal means

<code>ciWidthEmm</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
<code>emmPlots</code>	TRUE (default) or FALSE, provide estimated marginal means plots
<code>emmTables</code>	TRUE or FALSE (default), provide estimated marginal means tables
<code>emmWeights</code>	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency
<code>class</code>	TRUE or FALSE (default), provide a predicted classification table (or confusion matrix)
<code>acc</code>	TRUE or FALSE (default), provide the predicted accuracy of outcomes grouped by the cut-off value
<code>spec</code>	TRUE or FALSE (default), provide the predicted specificity of outcomes grouped by the cut-off value
<code>sens</code>	TRUE or FALSE (default), provide the predicted sensitivity of outcomes grouped by the cut-off value
<code>auc</code>	TRUE or FALSE (default), provide the rea under the ROC curve (AUC)
<code>rocPlot</code>	TRUE or FALSE (default), provide a ROC curve plot
<code>cutOff</code>	TRUE or FALSE (default), set a cut-off used for the predictions
<code>cutOffPlot</code>	TRUE or FALSE (default), provide a cut-off plot
<code>collin</code>	TRUE or FALSE (default), provide VIF and tolerance collinearity statistics
<code>boxTidwell</code>	TRUE or FALSE (default), provide Box-Tidwell test for linearity of the logit
<code>cooks</code>	TRUE or FALSE (default), provide summary statistics for the Cook's distance

### Value

A results object containing:

<code>results\$modelFit</code>	a table
<code>results\$modelComp</code>	a table
<code>results\$models</code>	an array of model specific results

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$modelFit$asDF
as.data.frame(results$modelFit)
```

### Examples

```
data('birthwt', package='MASS')

dat <- data.frame(
  low = factor(birthwt$low),
  age = birthwt$age,
  bwt = birthwt$bwt)
```

```

logRegBin(data = dat, dep = low,
          covs = vars(age, bwt),
          blocks = list(list("age", "bwt")),
          refLevels = list(list(var="low", ref="0")))

#
#  BINOMIAL LOGISTIC REGRESSION
#
#  Model Fit Measures
#  -----
#      Model    Deviance    AIC    R2-McF
#  -----
#      1     4.97e-7     6.00    1.000
#  -----
#
#
#  MODEL SPECIFIC RESULTS
#
#  MODEL 1
#
#  Model Coefficients
#  -----
#      Predictor    Estimate     SE      Z      p
#  -----
#      Intercept   2974.73225  218237.2   0.0136  0.989
#      age        -0.00653    482.7    -1.35e-5  1.000
#      bwt        -1.18532    87.0     -0.0136  0.989
#  -----
#      Note. Estimates represent the log odds of "low = 1"
#            vs. "low = 0"
#
#

```

**logRegMulti***Multinomial Logistic Regression***Description**

Multinomial Logistic Regression

**Usage**

```
logRegMulti(
  data,
  dep,
  covs = NULL,
  factors = NULL,
  blocks = list(list()),
  refLevels = NULL,
```

```

modelTest = FALSE,
dev = TRUE,
aic = TRUE,
bic = FALSE,
pseudoR2 = list("r2mf"),
omni = FALSE,
ci = FALSE,
ciWidth = 95,
OR = FALSE,
ciOR = FALSE,
ciWidthOR = 95,
emMeans = list(list()),
ciEmm = TRUE,
ciWidthEmm = 95,
emmPlots = TRUE,
emmTables = FALSE,
emmWeights = TRUE
)

```

## Arguments

<code>data</code>	the data as a data frame
<code>dep</code>	a string naming the dependent variable from <code>data</code> , variable must be a factor
<code>covs</code>	a vector of strings naming the covariates from <code>data</code>
<code>factors</code>	a vector of strings naming the fixed factors from <code>data</code>
<code>blocks</code>	a list containing vectors of strings that name the predictors that are added to the model. The elements are added to the model according to their order in the list
<code>refLevels</code>	a list of lists specifying reference levels of the dependent variable and all the factors
<code>modelTest</code>	TRUE or FALSE (default), provide the model comparison between the models and the NULL model
<code>dev</code>	TRUE (default) or FALSE, provide the deviance (or -2LogLikelihood) for the models
<code>aic</code>	TRUE (default) or FALSE, provide Aikaike's Information Criterion (AIC) for the models
<code>bic</code>	TRUE or FALSE (default), provide Bayesian Information Criterion (BIC) for the models
<code>pseudoR2</code>	one or more of 'r2mf', 'r2cs', or 'r2n'; use McFadden's, Cox & Snell, and Nagelkerke pseudo-R <sup>2</sup> , respectively
<code>omni</code>	TRUE or FALSE (default), provide the omnibus likelihood ratio tests for the predictors
<code>ci</code>	TRUE or FALSE (default), provide a confidence interval for the model coefficient estimates
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width

OR	TRUE or FALSE (default), provide the exponential of the log-odds ratio estimate, or the odds ratio estimate
ciOR	TRUE or FALSE (default), provide a confidence interval for the model coefficient odds ratio estimates
ciWidthOR	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
emMeans	a list of lists specifying the variables for which the estimated marginal means need to be calculate. Supports up to three variables per term.
ciEmm	TRUE (default) or FALSE, provide a confidence interval for the estimated marginal means
ciWidthEmm	a number between 50 and 99.9 (default: 95) specifying the confidence interval width for the estimated marginal means
emmPlots	TRUE (default) or FALSE, provide estimated marginal means plots
emmTables	TRUE or FALSE (default), provide estimated marginal means tables
emmWeights	TRUE (default) or FALSE, weigh each cell equally or weigh them according to the cell frequency

## Value

A results object containing:

results\$modelFit	a table
results\$modelComp	a table
results\$models	an array of model specific results

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$modelFit$asDF
as.data.frame(results$modelFit)
```

## Examples

```
data('birthwt', package='MASS')

dat <- data.frame(
  race = factor(birthwt$race),
  age = birthwt$age,
  low = factor(birthwt$low))

logRegMulti(data = dat, dep = race,
            covs = age, factors = low,
            blocks = list(list("age", "low")),
            refLevels = list(
              list(var="race", ref="1"),
              list(var="low", ref="0")))

#
```

```

# MULTINOMIAL LOGISTIC REGRESSION
#
# Model Fit Measures
# -----
#   Model Deviance AIC R2-McF
# -----
#   1      360    372  0.0333
# -----
#
#
# MODEL SPECIFIC RESULTS
#
# MODEL 1
#
# Model Coefficients
# -----
#   race Predictor Estimate SE Z p
# -----
#   2 - 1 Intercept 0.8155 1.1186 0.729 0.466
#   age       -0.1038 0.0487 -2.131 0.033
#   low:
#   1 0       0.7527 0.4700 1.601 0.109
#   3 - 1 Intercept 1.0123 0.7798 1.298 0.194
#   age       -0.0663 0.0324 -2.047 0.041
#   low:
#   1 0       0.5677 0.3522 1.612 0.107
# -----
#
#

```

**Description**

Ordinal Logistic Regression

**Usage**

```
logRegOrd(
  data,
  dep,
  covs = NULL,
  factors = NULL,
  blocks = list(list()),
  refLevels = NULL,
  modelTest = FALSE,
  dev = TRUE,
```

```

    aic = TRUE,
    bic = FALSE,
    pseudoR2 = list("r2mf"),
    omni = FALSE,
    thres = FALSE,
    ci = FALSE,
    ciWidth = 95,
    OR = FALSE,
    ciOR = FALSE,
    ciWidthOR = 95
)

```

## Arguments

<code>data</code>	the data as a data frame
<code>dep</code>	a string naming the dependent variable from <code>data</code> , variable must be a factor
<code>covs</code>	a vector of strings naming the covariates from <code>data</code>
<code>factors</code>	a vector of strings naming the fixed factors from <code>data</code>
<code>blocks</code>	a list containing vectors of strings that name the predictors that are added to the model. The elements are added to the model according to their order in the list
<code>refLevels</code>	a list of lists specifying reference levels of the dependent variable and all the factors
<code>modelTest</code>	TRUE or FALSE (default), provide the model comparison between the models and the NULL model
<code>dev</code>	TRUE (default) or FALSE, provide the deviance (or -2LogLikelihood) for the models
<code>aic</code>	TRUE (default) or FALSE, provide Aikaike's Information Criterion (AIC) for the models
<code>bic</code>	TRUE or FALSE (default), provide Bayesian Information Criterion (BIC) for the models
<code>pseudoR2</code>	one or more of ' <code>r2mf</code> ', ' <code>r2cs</code> ', or ' <code>r2n</code> '; use McFadden's, Cox & Snell, and Nagelkerke pseudo-R <sup>2</sup> , respectively
<code>omni</code>	TRUE or FALSE (default), provide the omnibus likelihood ratio tests for the predictors
<code>thres</code>	TRUE or FALSE (default), provide the thresholds that are used as cut-off scores for the levels of the dependent variable
<code>ci</code>	TRUE or FALSE (default), provide a confidence interval for the model coefficient estimates
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width
<code>OR</code>	TRUE or FALSE (default), provide the exponential of the log-odds ratio estimate, or the odds ratio estimate
<code>ciOR</code>	TRUE or FALSE (default), provide a confidence interval for the model coefficient odds ratio estimates
<code>ciWidthOR</code>	a number between 50 and 99.9 (default: 95) specifying the confidence interval width

**Value**

A results object containing:

<code>results\$modelFit</code>	a table
<code>results\$modelComp</code>	a table
<code>results\$models</code>	an array of model specific results

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$modelFit$asDF
as.data.frame(results$modelFit)
```

**Examples**

```
set.seed(1337)

y <- factor(sample(1:3, 100, replace = TRUE))
x1 <- rnorm(100)
x2 <- rnorm(100)

df <- data.frame(y=y, x1=x1, x2=x2)

logRegOrd(data = df, dep = y,
           covs = vars(x1, x2),
           blocks = list(list("x1", "x2")))

#
# ORDINAL LOGISTIC REGRESSION
#
# Model Fit Measures
# -----
#   Model    Deviance    AIC    R²-McF
# -----
#     1        218       226    5.68e-4
# -----
#
#
# MODEL SPECIFIC RESULTS
#
# MODEL 1
#
# Model Coefficients
# -----
#   Predictor    Estimate      SE       Z       p
# -----
#     x1          0.0579    0.193    0.300   0.764
#     x2          0.0330    0.172    0.192   0.848
# -----
# 
```

---

mancovaMANCOVA

---

### Description

Multivariate Analysis of (Co)Variance (MANCOVA) is used to explore the relationship between multiple dependent variables, and one or more categorical and/or continuous explanatory variables.

### Usage

```
mancova(
  data,
  deps,
  factors = NULL,
  covs = NULL,
  multivar = list("pillai", "wilks", "hotel", "roy"),
  boxM = FALSE,
  shapiro = FALSE,
  qqPlot = FALSE
)
```

### Arguments

<code>data</code>	the data as a data frame
<code>deps</code>	a string naming the dependent variable from <code>data</code> , variable must be numeric
<code>factors</code>	a vector of strings naming the factors from <code>data</code>
<code>covs</code>	a vector of strings naming the covariates from <code>data</code>
<code>multivar</code>	one or more of 'pillai', 'wilks', 'hotel', or 'roy'; use Pillai's Trace, Wilks' Lambda, Hotelling's Trace, and Roy's Largest Root multivariate statistics, respectively
<code>boxM</code>	TRUE or FALSE (default), provide Box's M test
<code>shapiro</code>	TRUE or FALSE (default), provide Shapiro-Wilk test
<code>qqPlot</code>	TRUE or FALSE (default), provide a Q-Q plot of multivariate normality

### Value

A results object containing:

<code>results\$multivar</code>	a table
<code>results\$univar</code>	a table
<code>results\$assump\$boxM</code>	a table
<code>results\$assump\$shapiro</code>	a table
<code>results\$assump\$qqPlot</code>	an image

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$multivar$asDF
as.data.frame(results$multivar)
```

## Examples

```
data('iris')

mancova(data = iris,
         deps = vars(Sepal.Length, Sepal.Width, Petal.Length, Petal.Width),
         factors = Species)

#
# MANCOVA
#
# Multivariate Tests
# -----
#          value   F    df1   df2   p
# -----
#   Species   Pillai's Trace   1.19  53.5     8   290 < .001
#           Wilks' Lambda  0.0234   199     8   288 < .001
#           Hotelling's Trace 32.5   581     8   286 < .001
#           Roy's Largest Root 32.2  1167     4   145 < .001
# -----
# 
# 
# Univariate Tests
# -----
#          Dependent Variable   Sum of Squares   df   Mean Square   F   p
# -----
#   Species   Sepal.Length      63.21       2   31.6061   119.3 < .001
#           Sepal.Width       11.34       2    5.6725    49.2 < .001
#           Petal.Length      437.10      2   218.5514  1180.2 < .001
#           Petal.Width       80.41       2   40.2067   960.0 < .001
#   Residuals   Sepal.Length     38.96     147    0.2650
#           Sepal.Width       16.96     147    0.1154
#           Petal.Length      27.22     147    0.1852
#           Petal.Width        6.16     147    0.0419
# -----
# 
```

## Description

Principal Component Analysis

**Usage**

```
pca(  
  data,  
  vars,  
  nFactorMethod = "parallel",  
  nFactors = 1,  
  minEigen = 1,  
  rotation = "varimax",  
  hideLoadings = 0.3,  
  sortLoadings = FALSE,  
  screePlot = FALSE,  
  eigen = FALSE,  
  factorCor = FALSE,  
  factorSummary = FALSE,  
  kmo = FALSE,  
  bartlett = FALSE  
)
```

**Arguments**

data	the data as a data frame
vars	a vector of strings naming the variables of interest in data
nFactorMethod	'parallel' (default), 'eigen' or 'fixed', the way to determine the number of factors
nFactors	an integer (default: 1), the number of components in the model
minEigen	a number (default: 1), the minimal eigenvalue for a component to be included in the model
rotation	'none', 'varimax' (default), 'quartimax', 'promax', 'oblimin', or 'simplimax', the rotation to use in estimation
hideLoadings	a number (default: 0.3), hide loadings below this value
sortLoadings	TRUE or FALSE (default), sort the factor loadings by size
screePlot	TRUE or FALSE (default), show scree plot
eigen	TRUE or FALSE (default), show eigenvalue table
factorCor	TRUE or FALSE (default), show factor correlations
factorSummary	TRUE or FALSE (default), show factor summary
kmo	TRUE or FALSE (default), show Kaiser-Meyer-Olkin (KMO) measure of sampling adequacy (MSA) results
bartlett	TRUE or FALSE (default), show Bartlett's test of sphericity results

**Value**

A results object containing:

results\$loadings

a table

results\$factorStats\$factorSummary	a table
results\$factorStats\$factorCor	a table
results\$modelFit\$fit	a table
results\$assump\$bartlett	a table
results\$assump\$kmo	a table
results\$eigen\$initEigen	a table
results\$eigen\$screePlot	an image

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$loadings$asDF
as.data.frame(results$loadings)
```

## Examples

```
data('iris')

pca(iris, vars = vars(Sepal.Length, Sepal.Width, Petal.Length, Petal.Width))

#
# PRINCIPAL COMPONENT ANALYSIS
#
# Component Loadings
# -----
#          1      Uniqueness
# -----
#   Sepal.Length    0.890    0.2076
#   Sepal.Width     -0.460   0.7883
#   Petal.Length    0.992    0.0168
#   Petal.Width     0.965    0.0688
# -----
#   Note. 'varimax' rotation was used
#
```

## Description

The Binomial test is used to test the Null hypothesis that the proportion of observations match some expected value. If the p-value is low, this suggests that the Null hypothesis is false, and that the true proportion must be some other value.

**Usage**

```
propTest2(
  data,
  vars,
  areCounts = FALSE,
  testValue = 0.5,
  hypothesis = "notequal",
  ci = FALSE,
  ciWidth = 95,
  bf = FALSE,
  priorA = 1,
  priorB = 1,
  ciBayes = FALSE,
  ciBayesWidth = 95,
  postPlots = FALSE
)
```

**Arguments**

<code>data</code>	the data as a data frame
<code>vars</code>	a vector of strings naming the variables of interest in <code>data</code>
<code>areCounts</code>	TRUE or FALSE (default), the variables are counts
<code>testValue</code>	a number (default: 0.5), the value for the null hypothesis
<code>hypothesis</code>	'notequal' (default), 'greater' or 'less', the alternative hypothesis
<code>ci</code>	TRUE or FALSE (default), provide confidence intervals
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95), the confidence interval width
<code>bf</code>	TRUE or FALSE (default), provide Bayes factors
<code>priorA</code>	a number (default: 1), the beta prior 'a' parameter
<code>priorB</code>	a number (default: 1), the beta prior 'b' parameter
<code>ciBayes</code>	TRUE or FALSE (default), provide Bayesian credible intervals
<code>ciBayesWidth</code>	a number between 50 and 99.9 (default: 95), the credible interval width
<code>postPlots</code>	TRUE or FALSE (default), provide posterior plots

**Value**

A results object containing:

<code>results\$table</code>	a table of the proportions and test results
<code>results\$postPlots</code>	an array of the posterior plots

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$table$asDF
as.data.frame(results$table)
```

## Examples

```
## Not run:
dat <- data.frame(x=c(8, 15))

propTest2(dat, vars = x, areCounts = TRUE)

#
# PROPORTION TEST (2 OUTCOMES)
#
# Binomial Test
# -----
#      Level   Count   Total   Proportion    p
# -----
#      x     1       8      23      0.348    0.210
#      2     15      23      0.652    0.210
# -----
# Note. Ha is proportion != 0.5
#
## End(Not run)
```

**propTestN**

*Proportion Test (N Outcomes)*

## Description

The  $\chi^2$  Goodness of fit test (not to be confused with the  $\chi^2$  test of independence), tests the Null hypothesis that the proportions of observations match some expected proportions. If the p-value is low, this suggests that the Null hypothesis is false, and that the true proportions are different to those tested.

## Usage

```
propTestN(data, var, counts = NULL, expected = FALSE, ratio = NULL, formula)
```

## Arguments

<code>data</code>	the data as a data frame
<code>var</code>	the variable of interest in data (not necessary when using a formula, see the examples)
<code>counts</code>	the counts in data
<code>expected</code>	TRUE or FALSE (default), whether expected counts should be displayed
<code>ratio</code>	a vector of numbers: the expected proportions
<code>formula</code>	(optional) the formula to use, see the examples

## Value

A results object containing:

<code>results\$props</code>	a table of the proportions
<code>results\$tests</code>	a table of the test results

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$props$asDF
as.data.frame(results$props)
```

## Examples

```
data('HairEyeColor')
dat <- as.data.frame(HairEyeColor)

propTestN(formula = Freq ~ Eye, data = dat, ratio = c(1,1,1,1))

#
# PROPORTION TEST (N OUTCOMES)
#
# Proportions
# -----
#   Level   Count   Proportion
# -----
#   Brown    220     0.372
#   Blue     215     0.363
#   Hazel    93      0.157
#   Green    64      0.108
# -----
#
#
# X2 Goodness of Fit
# -----
#   X2   df   p
# -----
#   133    3   < .001
# -----
```

## Description

Reliability Analysis

**Usage**

```
reliability(
  data,
  vars,
  alphaScale = TRUE,
  omegaScale = FALSE,
  meanScale = FALSE,
  sdScale = FALSE,
  corPlot = FALSE,
  alphaItems = FALSE,
  omegaItems = FALSE,
  meanItems = FALSE,
  sdItems = FALSE,
  itemRestCor = FALSE,
  revItems = NULL
)
```

**Arguments**

<code>data</code>	the data as a data frame
<code>vars</code>	a vector of strings naming the variables of interest in <code>data</code>
<code>alphaScale</code>	TRUE (default) or FALSE, provide Cronbach's alpha
<code>omegaScale</code>	TRUE or FALSE (default), provide McDonald's omega
<code>meanScale</code>	TRUE or FALSE (default), provide the mean
<code>sdScale</code>	TRUE or FALSE (default), provide the standard deviation
<code>corPlot</code>	TRUE or FALSE (default), provide a correlation plot
<code>alphaItems</code>	TRUE or FALSE (default), provide what the Cronbach's alpha would be if the item was dropped
<code>omegaItems</code>	TRUE or FALSE (default), provide what the McDonald's omega would be if the item was dropped
<code>meanItems</code>	TRUE or FALSE (default), provide item means
<code>sdItems</code>	TRUE or FALSE (default), provide item standard deviations
<code>itemRestCor</code>	TRUE or FALSE (default), provide item-rest correlations
<code>revItems</code>	a vector containing strings naming the varibales that are reverse scaled

**Value**

A results object containing:

<code>results\$scale</code>	a table
<code>results\$items</code>	a table
<code>results\$corPlot</code>	an image

Tables can be converted to data frames with `asDF` or `as.data.frame`. For example:

```
results$scale$asDF  
as.data.frame(results$scale)
```

## Examples

```
data('iris')  
  
reliability(iris, vars = c('Sepal.Length', 'Sepal.Width', 'Petal.Length', 'Petal.Width'),  
            omegaScale = TRUE)  
  
#  
# RELIABILITY ANALYSIS  
#  
# Scale Reliability Statistics  
# -----  
# Cronbach's alpha McDonald's omega  
# -----  
# scale 0.708 0.848  
# -----  
#
```

---

ttestIS

*Independent Samples T-Test*

---

## Description

The Student's Independent samples t-test (sometimes called a two-samples t-test) is used to test the null hypothesis that two groups have the same mean. A low p-value suggests that the null hypothesis is not true, and therefore the group means are different.

## Usage

```
ttestIS(  
  data,  
  vars,  
  group,  
  students = TRUE,  
  bf = FALSE,  
  bfPrior = 0.707,  
  welchs = FALSE,  
  mann = FALSE,  
  hypothesis = "different",  
  norm = FALSE,  
  qq = FALSE,  
  eqv = FALSE,  
  meanDiff = FALSE,
```

```

  ci = FALSE,
  ciWidth = 95,
  effectSize = FALSE,
  ciES = FALSE,
  ciWidthES = 95,
  desc = FALSE,
  plots = FALSE,
  miss = "perAnalysis",
  formula
)

```

## Arguments

<code>data</code>	the data as a data frame
<code>vars</code>	the dependent variables (not necessary when using a formula, see the examples)
<code>group</code>	the grouping variable with two levels (not necessary when using a formula, see the examples)
<code>students</code>	TRUE (default) or FALSE, perform Student's t-tests
<code>bf</code>	TRUE or FALSE (default), provide Bayes factors
<code>bfPrior</code>	a number between 0.5 and 2 (default 0.707), the prior width to use in calculating Bayes factors
<code>welchs</code>	TRUE or FALSE (default), perform Welch's t-tests
<code>mann</code>	TRUE or FALSE (default), perform Mann-Whitney U tests
<code>hypothesis</code>	'different' (default), 'oneGreater' or 'twoGreater', the alternative hypothesis; group 1 different to group 2, group 1 greater than group 2, and group 2 greater than group 1 respectively
<code>norm</code>	TRUE or FALSE (default), perform Shapiro-Wilk tests of normality
<code>qq</code>	TRUE or FALSE (default), provide Q-Q plots of residuals
<code>eqv</code>	TRUE or FALSE (default), perform Levene's tests for homogeneity of variances
<code>meanDiff</code>	TRUE or FALSE (default), provide means and standard errors
<code>ci</code>	TRUE or FALSE (default), provide confidence intervals
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95), the width of confidence intervals
<code>effectSize</code>	TRUE or FALSE (default), provide effect sizes
<code>ciES</code>	TRUE or FALSE (default), provide confidence intervals for the effect-sizes
<code>ciWidthES</code>	a number between 50 and 99.9 (default: 95), the width of confidence intervals for the effect sizes
<code>desc</code>	TRUE or FALSE (default), provide descriptive statistics
<code>plots</code>	TRUE or FALSE (default), provide descriptive plots
<code>miss</code>	'perAnalysis' or 'listwise', how to handle missing values; 'perAnalysis' excludes missing values for individual dependent variables, 'listwise' excludes a row from all analyses if one of its entries is missing.
<code>formula</code>	(optional) the formula to use, see the examples

## Details

The Student's independent t-test assumes that the data from each group are from a normal distribution, and that the variances of these groups are equal. If unwilling to assume the groups have equal variances, the Welch's t-test can be used in it's place. If one is additionally unwilling to assume the data from each group are from a normal distribution, the non-parametric Mann-Whitney U test can be used instead (However, note that the Mann-Whitney U test has a slightly different null hypothesis; that the distributions of each group is equal).

## Value

A results object containing:

results\$ttest	a table containing the t-test results
results\$assum\$norm	a table containing the normality tests
results\$assum\$eqv	a table containing the homogeneity of variances tests
results\$desc	a table containing the group descriptives
results\$plots	an array of groups of plots

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$ttest$asDF
as.data.frame(results$ttest)
```

## Examples

```
data('ToothGrowth')
ttestIS(formula = len ~ supp, data = ToothGrowth)

#
# INDEPENDENT SAMPLES T-TEST
#
# Independent Samples T-Test
# -----
#               statistic      df      p
# -----
#   len   Student's t       1.92    58.0   0.060
# -----
```

## Description

The Student's One-sample t-test is used to test the null hypothesis that the true mean is equal to a particular value (typically zero). A low p-value suggests that the null hypothesis is not true, and therefore the true mean must be different from the test value.

**Usage**

```
ttestOneS(
  data,
  vars,
  students = TRUE,
  bf = FALSE,
  bfPrior = 0.707,
  wilcoxon = FALSE,
  testValue = 0,
  hypothesis = "dt",
  norm = FALSE,
  qq = FALSE,
  meanDiff = FALSE,
  ci = FALSE,
  ciWidth = 95,
  effectSize = FALSE,
  ciES = FALSE,
  ciWidthES = 95,
  desc = FALSE,
  plots = FALSE,
  miss = "perAnalysis",
  mann = FALSE
)
```

**Arguments**

<code>data</code>	the data as a data frame
<code>vars</code>	a vector of strings naming the variables of interest in <code>data</code>
<code>students</code>	TRUE (default) or FALSE, perform Student's t-tests
<code>bf</code>	TRUE or FALSE (default), provide Bayes factors
<code>bfPrior</code>	a number between 0.5 and 2.0 (default 0.707), the prior width to use in calculating Bayes factors
<code>wilcoxon</code>	TRUE or FALSE (default), perform Wilcoxon signed rank tests
<code>testValue</code>	a number specifying the value of the null hypothesis
<code>hypothesis</code>	' <code>dt</code> ' (default), ' <code>gt</code> ' or ' <code>lt</code> ', the alternative hypothesis; different to <code>testValue</code> , greater than <code>testValue</code> , and less than <code>testValue</code> respectively
<code>norm</code>	TRUE or FALSE (default), perform Shapiro-wilk tests of normality
<code>qq</code>	TRUE or FALSE (default), provide a Q-Q plot of residuals
<code>meanDiff</code>	TRUE or FALSE (default), provide means and standard deviations
<code>ci</code>	TRUE or FALSE (default), provide confidence intervals for the mean difference
<code>ciWidth</code>	a number between 50 and 99.9 (default: 95), the width of confidence intervals
<code>effectSize</code>	TRUE or FALSE (default), provide Cohen's d effect sizes
<code>ciES</code>	TRUE or FALSE (default), provide confidence intervals for the effect-sizes

ciWidthES	a number between 50 and 99.9 (default: 95), the width of confidence intervals for the effect sizes
desc	TRUE or FALSE (default), provide descriptive statistics
plots	TRUE or FALSE (default), provide descriptive plots
miss	'perAnalysis' or 'listwise', how to handle missing values; 'perAnalysis' excludes missing values for individual dependent variables, 'listwise' excludes a row from all analyses if one of its entries is missing.
mann	deprecated

## Details

The Student's One-sample t-test assumes that the data are from a normal distribution – in the case that one is unwilling to assume this, the non-parametric Wilcoxon signed-rank can be used in it's place (However, note that the Wilcoxon signed-rank has a slightly different null hypothesis; that the \*median\* is equal to the test value).

## Value

A results object containing:

results\$ttest	a table containing the t-test results
results\$normality	a table containing the normality test results
results\$descriptives	a table containing the descriptives
results\$plots	an image of the descriptive plots
results\$qq	an array of Q-Q plots

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$ttest$asDF
as.data.frame(results$ttest)
```

## Examples

```
data('ToothGrowth')

ttestOneS(ToothGrowth, vars = vars(len, dose))

#
# ONE SAMPLE T-TEST
#
# One Sample T-Test
# -----
#               statistic   df      p
# -----
#   len     Student's t      19.1  59.0    < .001
#   dose    Student's t      14.4  59.0    < .001
# -----
```

---

ttestPS*Paired Samples T-Test*

---

**Description**

The Student's paired samples t-test (sometimes called a dependent-samples t-test) is used to test the null hypothesis that the difference between pairs of measurements is equal to zero. A low p-value suggests that the null hypothesis is not true, and that the difference between the measurement pairs is not zero.

**Usage**

```
ttestPS(
  data,
  pairs,
  students = TRUE,
  bf = FALSE,
  bfPrior = 0.707,
  wilcoxon = FALSE,
  hypothesis = "different",
  norm = FALSE,
  qq = FALSE,
  meanDiff = FALSE,
  ci = FALSE,
  ciWidth = 95,
  effectSize = FALSE,
  ciES = FALSE,
  ciWidthES = 95,
  desc = FALSE,
  plots = FALSE,
  miss = "perAnalysis"
)
```

**Arguments**

<code>data</code>	the data as a data frame
<code>pairs</code>	a list of lists specifying the pairs of measurement in <code>data</code>
<code>students</code>	TRUE (default) or FALSE, perform Student's t-tests
<code>bf</code>	TRUE or FALSE (default), provide Bayes factors
<code>bfPrior</code>	a number between 0.5 and 2 (default 0.707), the prior width to use in calculating Bayes factors
<code>wilcoxon</code>	TRUE or FALSE (default), perform Wilcoxon signed rank tests
<code>hypothesis</code>	'different' (default), 'oneGreater' or 'twoGreater', the alternative hypothesis; measure 1 different to measure 2, measure 1 greater than measure 2, and measure 2 greater than measure 1 respectively

norm	TRUE or FALSE (default), perform Shapiro-wilk normality tests
qq	TRUE or FALSE (default), provide a Q-Q plot of residuals
meanDiff	TRUE or FALSE (default), provide means and standard errors
ci	TRUE or FALSE (default), provide confidence intervals
ciWidth	a number between 50 and 99.9 (default: 95), the width of confidence intervals
effectSize	TRUE or FALSE (default), provide effect sizes
cIES	TRUE or FALSE (default), provide confidence intervals for the effect-sizes
ciWidthES	a number between 50 and 99.9 (default: 95), the width of confidence intervals for the effect sizes
desc	TRUE or FALSE (default), provide descriptive statistics
plots	TRUE or FALSE (default), provide descriptive plots
miss	'perAnalysis' or 'listwise', how to handle missing values; 'perAnalysis' excludes missing values for individual dependent variables, 'listwise' excludes a row from all analyses if one of its entries is missing

## Details

The Student's paired samples t-test assumes that pair differences follow a normal distribution – in the case that one is unwilling to assume this, the non-parametric Wilcoxon signed-rank can be used in its place (However, note that the Wilcoxon signed-rank has a slightly different null hypothesis; that the two groups of measurements follow the same distribution).

## Value

A results object containing:

results\$ttest	a table containing the t-test results
results\$norm	a table containing the normality test results
results\$desc	a table containing the descriptives
results\$plots	an array of the descriptive plots

Tables can be converted to data frames with `asDF` or [as.data.frame](#). For example:

```
results$ttest$asDF
as.data.frame(results$ttest)
```

## Examples

```
## Not run:
data('bugs', package = 'jmv')

ttestPS(bugs, pairs = list(
  list(i1 = 'LDLF', i2 = 'LDHF')))

#
# PAIRED SAMPLES T-TEST
```

```
#  
# Paired Samples T-Test  
# -----  
#  
#          statistic    df     p  
# -----  
#   LDLF    LDHF  Student's t      -6.65    90.0  < .001  
# -----  
#  
## End(Not run)
```

# Index

ancova, 2  
ANOVA, 5  
anovaNP, 7  
anovaOneW, 9  
anovaRM, 11  
anovaRMNP, 14  
as.data.frame, 4, 7, 8, 10, 13, 15, 18, 21, 23,  
25, 28, 33, 37, 40, 43, 46, 48, 50, 51,  
53, 55, 57, 59, 61  
bugs, 15  
cfa, 16  
contTables, 19  
contTablesPaired, 22  
corrMatrix, 23  
descriptives, 26  
efa, 29  
linReg, 31  
logLinear, 34  
logRegBin, 38  
logRegMulti, 41  
logRegOrd, 44  
mancova, 47  
pca, 48  
propTest2, 50  
propTestN, 52  
reliability, 53  
ttestIS, 55  
ttestOneS, 57  
ttestPS, 60