

# Package ‘ampir’

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**Type** Package

**Title** Predict Antimicrobial Peptides

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## Description

A toolkit to predict antimicrobial peptides from protein sequences on a genome-wide scale. It incorporates two support vector machine models ("precursor" and "mature") trained on publicly available antimicrobial peptide data using calculated physico-chemical and compositional sequence properties described in Meher et al. (2017) <doi:10.1038/srep42362>. In order to support genome-wide analyses, these models are designed to accept any type of protein as input and calculation of compositional properties has been optimised for high-throughput use. For details see Fingerhut et al. 2020 <doi:10.1101/2020.05.07.082412>.

**URL** <https://github.com/Legana/ampir>

**License** GPL-2

**Encoding** UTF-8

**LazyData** true

**Depends** R (>= 3.5.0)

**Imports** Peptides, caret (>= 6.0.0), kernlab, Rcpp, parallel

**RoxygenNote** 7.1.0

**Suggests** testthat, knitr, rmarkdown, e1071

**VignetteBuilder** knitr

**LinkingTo** Rcpp

**NeedsCompilation** yes

**Author** Legana Fingerhut [aut, cre] (<<https://orcid.org/0000-0002-2482-5336>>),

Ira Cooke [aut] (<<https://orcid.org/0000-0001-6520-1397>>),

Jinlong Zhang [ctb] (R/read\_faa.R),

Nan Xiao [ctb] (R/calc\_pseudo\_comp.R)

**Maintainer** Legana Fingerhut <[legana.fingerhut@my.jcu.edu.au](mailto:legana.fingerhut@my.jcu.edu.au)>

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<i>aaseq_is_valid</i>	<i>Check protein sequences for non-standard amino acids</i>
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### Description

Any proteins that contains an amino acid that is not one of the 20 standard amino acids is flagged as invalid

### Usage

```
aaseq_is_valid(seq)
```

### Arguments

seq	A vector of protein sequences
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### Value

A logical vector where TRUE indicates a valid protein sequence and FALSE indicates a sequence with invalid amino acids

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calculate\_features     *Calculate a set of numerical features from protein sequences*

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## Description

This function calculates set physicochemical and compositional features from protein sequences in preparation for supervised model learning

## Usage

```
calculate_features(df, min_len = 10)
```

## Arguments

df	A data frame which contains protein sequence names as the first column and amino acid sequence as the second column
min_len	Minimum length sequence for which features can be calculated. It is an error to provide sequences with length shorter than this

## Value

A data frame containing numerical values related to the protein features of each given protein

## Note

This function depends on the Peptides package

## References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015).

## Examples

```
my_protein_df <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

calculate_features(my_protein_df)
## Output (showing the first six output columns)
#   seq_name Amphiphilicity Hydrophobicity pI      Mw      Charge ....
# [1] G1P6H5_MYOLU    0.4145847     0.4373494  8.501312 9013.757  4.53015 ....
```

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**calc\_amphiphilicity**    *Calculate amphiphilicity (or hydrophobic moment)*

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### Description

Calculate amphiphilicity (or hydrophobic moment)

### Usage

`calc_amphiphilicity(seq)`

### Arguments

`seq`                  A protein sequence

### References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

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**calc\_hydrophobicity**    *Calculate the hydrophobicity*

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### Description

Calculate the hydrophobicity

### Usage

`calc_hydrophobicity(seq)`

### Arguments

`seq`                  A protein sequence

### References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

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`calc_mw`

*Calculate the molecular weight*

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### Description

Calculate the molecular weight

### Usage

`calc_mw(seq)`

### Arguments

`seq` A protein sequence

### References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

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`calc_net_charge`

*Calculate the net charge*

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### Description

Calculate the net charge

### Usage

`calc_net_charge(seq)`

### Arguments

`seq` A protein sequence

### References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

**calc\_pI***Calculate the isoelectric point (pI)***Description**

Calculate the isoelectric point (pI)

**Usage**

```
calc_pI(seq)
```

**Arguments**

seq	pI
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**References**

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal.* 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

**calc\_pseudo\_comp***Calculate the pseudo amino acid composition***Description**

This function is adapted from the extractPAAC function from the protr package (<https://github.com/nanxstats/protr>)

**Usage**

```
calc_pseudo_comp(seq, lambda_min = 4, lambda_max = 19)
```

**Arguments**

seq	A vector of protein sequences as character strings
lambda_min	Minimum allowable lambda. It is an error to provide a protein sequence shorter than lambda_min+1
lambda_max	For each sequence lambda will be set to one less than the sequence length or lambda_max, whichever is smaller

**References**

Nan Xiao, Dong-Sheng Cao, Min-Feng Zhu, and Qing-Song Xu. (2015). protr/ProtrWeb: R package and web server for generating various numerical representation schemes of protein sequences. *Bioinformatics* 31 (11), 1857-1859.

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chunk_rows	<i>Determine row breakpoints for dividing a dataset into chunks for parallel processing</i>
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**Description**

Determine row breakpoints for dividing a dataset into chunks for parallel processing

**Usage**

```
chunk_rows(nrows, n_cores)
```

**Arguments**

nrows	The number of rows in the dataset to be chunked
n_cores	The number of cores that will be used for parallel processing

**Value**

A list of integer vectors consisting of the rows in each chunk

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df_to_faa	<i>Save a dataframe in FASTA format</i>
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**Description**

This function writes a dataframe out as a FASTA format file

**Usage**

```
df_to_faa(df, file = "")
```

**Arguments**

df	a dataframe containing two columns: the sequence name and amino acid sequence itself
file	file path to save the named file to

**Value**

A FASTA file where protein sequences are represented in two lines: The protein name preceded by a greater than symbol, and a new second line that contains the protein sequence

## Examples

```
my_protein <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

# Write a dataframe to a FASTA file
df_to_faa(my_protein, tempfile("my_protein.fasta", tempdir()))
```

**predict\_amps**

*Predict the antimicrobial peptide probability of a protein*

## Description

This function predicts the probability of a protein to be an antimicrobial peptide

## Usage

```
predict_amps(faa_df, min_len = 5, n_cores = 1, model = "precursor")
```

## Arguments

<code>faa_df</code>	A dataframe obtained from <code>read_faa</code> containing two columns: the sequence name ( <code>seq_name</code> ) and amino acid sequence ( <code>seq_aa</code> )
<code>min_len</code>	The minimum protein length for which predictions will be generated
<code>n_cores</code>	On multicore machines split the task across this many processors. This option does not work on Windows
<code>model</code>	Either a string with the name of a built-in model ( <code>mature</code> , <code>precursor</code> ), OR, A train object suitable for passing to the <code>predict.train</code> function in the <code>caret</code> package. If omitted the default model will be used.

## Value

The original input data.frame with a new column added called `prob_AMP` with the probability of that sequence to be an antimicrobial peptide. Any sequences that are too short or which contain invalid amino acids will have NA in this column

## Examples

```
my_bat_faa_df <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

predict_amps(my_bat_faa_df)
#      seq_name    prob_AMP
# [1] G1P6H5_MYOLU  0.9723796
```

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read_faa	<i>Read FASTA amino acids file into a dataframe</i>
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## Description

This function reads a FASTA amino acids file into a dataframe

## Usage

```
read_faa(file = NULL)
```

## Arguments

file            file path to the FASTA format file containing the protein sequences

## Value

Dataframe containing the sequence name (seq\_name) and sequence (seq\_aa) columns

## Note

This function was adapted from ‘read.fasta.R’ by Jinlong Zhang (jinlongzhang01@gmail.com) for the phylotools package (<http://github.com/helixcn/phylotools>)

## Examples

```
read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

## Output
#      seq_name      seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAACLLLLLASLTSYSL....
```

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remove_nonstandard_aa	<i>Remove non standard amino acids from protein sequences</i>
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## Description

This function removes anything that is not one of the 20 standard amino acids in protein sequences

## Usage

```
remove_nonstandard_aa(df)
```

## Arguments

df            A dataframe which contains protein sequence names as the first column and amino acid sequence as the second column

**Value**

a dataframe like the input dataframe but with removed proteins that contained non standard amino acids

**Examples**

```
non_standard_df <- readRDS(system.file("extdata/non_standard_df.rds", package = "ampir"))

# non_standard_df
#   seq_name      seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAAACLLLLLALTSYSLLSQTTQLADLQTQ....
# [2] fake_sequence MKVTHEUSYR$GXMBIJIDG*M80-%

remove_nonstandard_aa(non_standard_df)
#   seq_name      seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAAACLLLLLALTSYSLLSQTTQLADLQTQ....
```

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