## Package 'XMRF'

June 25, 2015

Type Package
Title Markov Random Fields for High-Throughput Genetics Data
Version 1.0
Date 2015-06-12
Author
Ying-Wooi Wan, Genevera I. Allen, Yulia Baker, Eunho Yang, Pradeep Ravikumar, Zhandong Liu
Maintainer Ying-Wooi Wan <yingwoow@bcm.edu>
Depends R (>= 3.0.2)
Imports igraph, glmnet, MASS, snowfall, parallel, Matrix
Description Fit Markov Networks to a wide range of high-throughput genomics data.
License GPL-2
URL http://www.liuzlab.org/XMRF/
LazyLoad true

NeedsCompilation yes Repository CRAN Date/Publication 2015-06-25 07:07:40

## **R** topics documented:

MRF-package	2
cadat	4
gm.fit	4
MS	5
mbdaMax	6
ot.GMS	6
otGML	7
otNet	8
ocessSeq	9
MRF	10
MRF.Sim	12

14

Index

XMRF-package

#### Description

An R package to learn and visualize the underlying relationships between genes from various types of high-throughput genomics data.

#### Details

Package:	XMRF
Type:	Package
Version:	1.0
Date:	2015-06-12
License:	GPL-2

Technological advances have produced large amounts of high-throughput "omics" data that allow us to study the complicated interactions between genes, mutations, aberrations, and epi-genetic markers. Markov Random Fields (MRFs), or Markov Networks, enable us to estimate these genomics networks via sparse, high-dimensional undirected graphical models.

Here, we provide the community a convenient and useful tool to learn the complex genomics networks from various types of high-throughput genomics data. This package encodes the recently proposed parametric family of graphical models based on node-conditional univariate exponential family distributions (Yang *et. al*, 2012, 2013a). Specifically, our package has methods for estimating Gaussian graphical model (Meinshausen and Buhlmann, 2006), Ising model (Ravikumar *et. al*, 2010), and the Poisson family graphical models (Allen and Liu, 2012, 2013; Yang *et. al* 2013b). These models can be used to estimate genetic networks from a variety of data types:

Genomics Data	Туре	XMRF Family
RNA-Seq or miRNA-Seq	Counts	LPGM, SPGM
Microarray or Methylation array	Continuous	GGM
Mutation or CNVs	Binary	ISM

To estimate the network structures from different types of genomics data with this package, users simply need to specify the "method" in the main function, for example XMRF(..., method="LPGM") to fit LPGM to next-generation sequencing data.

In this package, we implement the neighborhood selection graph estimation technique by proximal or projected gradient descent using warm starts over the range of regularization parameters. This

technique allows estimation of the neighborhood for each node independently and thus can be done in parallel, thus speeding computation.

This package also implements two data-driven approaches to select the sparsity of a fitted network: a stability-based approach for a single regularization value over many bootstrap resamples (Mein-shausen and Buhlmann, 2010), or computed over a range of regularization values with StARS (Liu *et. al.*, 2010).

#### Author(s)

Ying-Wooi Wan, Genevara Allen, Yulia Baker, Eunho Yang, Pradeep Ravikumar, and Zhandong Liu

Maintainer: Ying-Wooi Wan<yingwoow@bcm.edu>

#### References

Allen, G.I., and Liu, Z. (2012). A Log-Linear graphical model for inferring genetic networks from high-throughput sequencing data. *The IEEE International Conference on Bioinformatics and Biomedicine (BIBM 2012)*.

Allen, G. I., and Liu, Z. (2013). A Local Poisson Graphical Model for Inferring Genetic Networks from Next Generation Sequencing Data. *IEEE Transactions on NanoBioscience*, **12**(3), pp.1-10

Liu, H., Roeder, K., and Wasserman, L. (2010). Stability approach to regularization selection (stars) for high dimensional graphical models. *NIPS 23*, pp.1432-1440.

Meinshausen, N. and Buhlmann, P. (2006). High-dimensional graphs and variable selection with the lasso. *The Annals of Statistics*, **34**(3), pp.1436-1462.

Meinshausen, N. and Buhlmann, P. (2010). Stability selection. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, **72**(4), pp.417-473.

Ravikumar, P., Wainwright, M., and Lafferty, J. (2010). High-dimensional ising model selection using 11-regularized logistic regression. *The Annals of Statistics*, **38**(3), pp.1287-1319.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2012). Graphical models via generalized linear models. *NIPS*, **25**, pp.1367–1375.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013a). On graphical models via univariate exponential family distributions. *arXiv preprint arXiv:1301.4183*.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013b). On Poisson graphical models. *NIPS*, pp.1718-1726.

#### See Also

XMRF

#### Examples

library(XMRF)

n = 100 p = 20

```
sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
simDat <- sim$X
# Compute the optimal lambda
lmax = lambdaMax(t(simDat))
lambda = 0.01* sqrt(log(p)/n) * lmax
# Run LPGM
lpgm.fit <- XMRF(simDat, method="LPGM", N=10, lambda.path=lambda, parallel=FALSE)
# Print the fitted Markov networks
lpgm.fit
ml = plotNet(sim$B)
ml = plot(lpgm.fit, mylayout=ml)</pre>
```

```
brcadat
```

4

RNA-Seq Data of BRCA Patients from TCGA

#### Description

A matrix of RNA-Seq read counts from BRCA patients.

#### Usage

data("brcadat")

#### Format

A matrix of level 3 RNA-Seq (UNC Illumina HiSeq RNASeqV2) data with 445 breast invasive carcinoma (BRCA) patients from the Cancer Genome Atlas (TCGA) project on 353 genes with somatic mutations listed in the Catalogue of Somatic Mutations in Cancer (COSMIC). The matrix is organized in dimension of *gene* x *sample*.

ggm.fit

Fitted Gaussian Graphical Models

#### Description

An example fitted Gaussian graphical model

#### Usage

data("ggm.fit")

#### Format

A GMS object.

#### GMS

#### Details

This fitted Gaussian graphical model is included as the example model for demonstrating the usage of other functions. The model was fitted with a simulated multivariate Gaussian data of scale-free graph structure, 30 nodes, and 200 observations. StARS stability selection of 100 iterations was used to identify the optimal network over the regularization path of 10 parameters.

#### See Also

GMS, plot.GMS

GMS

#### Graphical Models Object

#### Description

This class of objects is returned by the XMRF function in this package, to represent the fitted Markov Network(s). Objects of this class have the print method to display the core information of the fitted models and plot method to plot the optimal Markov Network.

#### Arguments

v	vector of (nlams length) network variability measured for each regularization level.
D	a list of <i>pxp</i> matrices of stability scores of inferred edges of each network along the regularization path.
lambda.path	numeric vector used for regularization path.
opt.lambda	lambda value that gives the optimal network (network with maximum variabil- ity).
network	a list of <i>pxp</i> matrices of fitted networks along the regularization path.
opt.index	index of the regularization value that gives the optimal network.

#### See Also

XMRF, plot.GMS

lambdaMax

#### Description

Compute the maximum lambda

#### Usage

lambdaMax(X)

## Arguments X

a *nxp* data matrix.

#### Details

Compute the largest value for regularization (maximum lambda) that gives the null model. The maximum lambda is computed based on the input data matrix, and is the maximum element from column-wise multiplication of data matrix normalized by the number of observations.

#### Value

a numeric value

plot.GMS

Plot GMS Object

#### Description

Default function to plot the network of a GMS object.

#### Usage

## S3 method for class 'GMS'
plot(x, fn = "", th = 1e-06, i = NULL, mylayout = NULL, vars = NULL, ...)

#### Arguments

х	a GMS object.
fn	file name to save the network plot; default to be an empty string, so the network is plotted to the standard output (screen). NOTE: if a file name is specified, it should be file name for PDF file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.

#### plotGML

i	index of the network (along the regularization path) to be plotted. Default NULL to plot the optimal network.	to
mylayout	graph layout to draw the network, default to NULL.	
vars	vector of variable names, default to NULL.	
	other generic arguments for plot method.	

#### Details

This is the default plotting function for GMS objects (Markov Networks inferred over a regularization path). Refer to GMS for details on GMS object. This function will plot the optimal network on the screen by default. However, given a file name, the plot will be saved to a PDF file. Also, given a specific index corresponding to the index of lambda.path, the associated network will be plotted.

The network will be plotted in force-directed layout (layout.fruchterman.reingold with default parameters implemented in igraph package).

#### Value

Returns the layout object from igraph package - numeric matrix of two columns and the rows with the same number as the number of vertices.

#### See Also

GMS

#### Examples

```
library(XMRF)
data('ggm.fit')
plot(ggm.fit, fn="ggm.fit.net.pdf")
```

plotGML

Plot Network in GML

#### Description

Plot the network in graph modeling language (GML).

#### Usage

```
plotGML(x, fn = "", th = 1e-06, i = NULL, weight = FALSE, vars = NULL)
```

#### Arguments

x	a GMS object.
fn	file name to save the GML file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.
i	index of the network (along the regularization path) to be plotted. Default to NULL for optimal network.
weight	boolean value to indicate if writing the stability on the inferred edges, default to FALSE.
vars	vector of variable names, default to NULL.

plotNet Plot Network
----------------------

#### Description

Plot a network with specific layout.

#### Usage

plotNet(net, fn = "", th = 1e-06, mylayout = NULL)

#### Arguments

net	a square adjacency matrix of the network to be plotted.
fn	file name to save the network plot. Default to be an empty string, so the network is plotted to the standard output (screen). NOTE: if a file name is specified, it should be file name for PDF file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.
mylayout	graph layout to draw the network, default to NULL.

#### Details

This function serves as the alternative plotting function to allow users to plot a specific network with specific layout, such as plotting the simulated network.

#### Value

Returns the layout object from igraph package - numeric matrix of two columns and the rows with the same number as the number of vertices.

#### processSeq

#### Examples

```
library(XMRF)
n = 200
p = 30
sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
ml = plotNet(sim$B)</pre>
```

```
processSeq
```

Process Sequencing Data for Poisson-based MRFs

#### Description

Process and normalize RNA-Sequencing count data into a distribution appropriate for Poisson MRFs.

#### Usage

processSeq(X, quanNorm = 0.75, nLowCount = 20, percentLowCount = 0.95, NumGenes = 500, PercentGenes = 0.1)

#### Arguments

Х	nxp data matrix.
quanNorm	an optional parameter controlling the quantile for sample normalization, default to 0.75.
nLowCount	minimum read count to decide if to filter a gene, default to 20.
percentLowCount	
	filter out a gene if it has this percentage of samples less than <code>nLowCount</code> , default to 0.95.
NumGenes	number of genes to retain in the final data set, default to 500.
PercentGenes	percentage of genes to retain, default to 0.1.

#### Details

To process the next-generation sequencing count data into proper distribution (with dispersion removed), the following steps are taken in this function:

- 1. Quantile normalization for the samples.
- 2. Filter out genes with all low counts.
- 3. Filter genes by maximal variance (if specified).
- 4. Transform the data to be closer to the Poisson distribution. A log or power transform is considered and selected based upon the Kolmogorov-Smirnov goodness of fit test.

#### Value

a *n* x NumGenes or PercentGenes processed data matrix.

#### Examples

```
library(XMRF)
data('brcadat')
brca = t(processSeq(t(brcadat), PercentGenes=1))
```

XMRF

Markov Random Fields for Exponential Family Distributions

#### Description

Infer networks from genomics data using Markov Random Fields specified by node-conditional univariate exponential family distributions.

#### Usage

```
XMRF(X, method = "LPGM", stability = "bootstrap", N = 100, beta = 0.01, lmin = 0.01,
nlams = 20, lambda.path = NULL, parallel = TRUE, nCpus = 4, sym = TRUE, th = 0.01,
sth = 0.95, R = max(X), R0 = 0)
```

#### Arguments

Х	a <i>p</i> x <i>n</i> data matrix.
method	specification of the type of MRF model, default to "LPGM" for log-linear Poisson- based graphical model. Other allowed methods are "PGM" for regular Poisson, "TPGM" for truncated Poisson, "SPGM" for sublinear Poisson, "GGM" for Gaussian graphical models, and "ISM" for Ising model.
stability	specification of the stability method, default to "bootstrap". Another accepted value is "star" for Stability Approach to Regularization Selection (StARS).
N	number of iterations for stability selection, default to 100.
beta	threshold value on sparsity of the network, default to 0.01.
lmin	ratio of minimum lambda value from the maximum lambda value, default to 0.01.
nlams	number of lambda for regularization, default to 20.
lambda.path	vector lambda used for regularization, default ot NULL.
parallel	logical value to indicate if the process should be run parallelly in multiple threads, default to TRUE.
nCpus	number of (maximum) cores to use for parallel execution, default to 4.
sym	logical value to indicate if symmetry is enforced on the inferred edges, default to TRUE.
th	threshold value for the estimated edge coefficient, default to 0.005.
sth	an inferred edge is retained only if its stability score is greater than sth, default to 0.9.
R	truncation level for classes "TPGM" and "SPGM". The value has to be positive. Default to the maximum value of the input data matrix.
RØ	lower-bound truncation level for "SPGM", default to 0.

10

#### XMRF

#### Details

This is the main function of the package that fits exponential family Markov Networks to genomics data. To estimate the network structures using native distribution of the genomics data, specify the MRF family types in the "method" parameter. For genomic networks based on next-generation sequencing data, we recommend using the LPGM family. The table at the beginning of the document lists the family type recommended for each of the genomic data platforms.

#### Value

An object of class GMS will be returned representing the inferred Markov networks over the regularization path. See GMS for details.

#### References

Allen, G.I., and Liu, Z. (2012). A Log-Linear graphical model for inferring genetic networks from high-throughput sequencing data. *The IEEE International Conference on Bioinformatics and Biomedicine (BIBM 2012)*.

Allen, G. I., and Liu, Z. (2013). A Local Poisson Graphical Model for Inferring Genetic Networks from Next Generation Sequencing Data. *IEEE Transactions on NanoBioscience*, **12**(3), pp.1-10

Liu, H., Roeder, K., and Wasserman, L. (2010). Stability approach to regularization selection (stars) for high dimensional graphical models. *NIPS 23*, pp.1432?1440.

Meinshausen, N. and Buhlmann, P. (2006). High-dimensional graphs and variable selection with the lasso. *The Annals of Statistics*, **34**(3), pp.1436?1462.

Meinshausen, N. and Buhlmann, P. (2010). Stability selection. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, **72**(4), pp.417?473.

Ravikumar, P., Wainwright, M., and Lafferty, J. (2010). High-dimensional ising model selection using 11-regularized logistic regression. *The Annals of Statistics*, **38**(3), pp.1287?1319.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2012). Graphical models via generalized linear models. *NIPS*, **25**, pp.1367–1375.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013a). On graphical models via univariate exponential family distributions. *arXiv preprint arXiv:1301.4183*.

Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013b). On Poisson graphical models. *NIPS*, pp.1718-1726.

#### See Also

XMRF-package, GMS, plot.GMS

#### Examples

```
# Example for LPGM
# Refer to the package's introduction for identical example
## Not run: n = 100
## Not run: p = 20
## Not run: sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
## Not run: simDat <- sim$X
## Not run: # Compute the optimal lambda</pre>
```

```
## Not run: lmax = lambdaMax(t(simDat))
## Not run: lambda = 0.01* sqrt(log(p)/n) * lmax
## Not run: # Run LPGM
## Not run: lpgm.fit <- XMRF(simDat, method="LPGM", N=10, lambda.path=lambda)
## Not run: ml = plotNet(sim$B, fn="simDat.netPlot.pdf")
## Not run: ml = plot(lpgm.fit, fn="lpgm.netPlot_1.pdf", i=1, mylayout=ml)
## Not run: plot(lpgm.fit, fn="lpgm.fit.net.pdf")</pre>
```

XMRF.Sim

Generate simulated data from XMRF models

#### Description

Generate data from different multivariate distributions with different network structures.

#### Usage

XMRF.Sim(n = 100, p = 50, model = "LPGM", graph.type = "scale-free")

#### Arguments

n	number of samples, default to 100.
р	number of variables, default to 50.
model	Markov Network models to indicate the distribution family of the data to be generated, default to "LPGM". Other model options include "PGM", "TPGM", "SPGM", "GGM" and "ISM".
graph.type	graph structure with 3 options: "scale-free", "hub", and "lattice". Default to "scale-free".

#### Details

This function will first generate a graph of the specified graph structure; then based on the generated network, it simulates a multivariate data matrix that follows distribution for the Markov Random Fields model specified.

#### Value

A list of two elements:

- B *pxp* adjacency matrix of the generated graph.
- X *pxn* data matrix.

#### XMRF.Sim

#### Examples

library(XMRF)

```
# simulate scale-free network and data of multivariate Poisson for LPGM
sim <- XMRF.Sim(n=100, p=20, model="LPGM", graph.type="scale-free")
hist(sim$X)
plotNet(sim$B)
# simulate hub network and data of multivariate Gaussian for GGM
sim <- XMRF.Sim(n=100, p=20, model="GGM", graph.type="hub")
hist(sim$X)
plotNet(sim$B)
# simulate hub network and data of multivariate bionomial for ISM
sim <- XMRF.Sim(n=100, p=15, model="ISM", graph.type="hub")
hist(sim$X)
plotNet(sim$B)
```

# Index

\*Topic datasets brcadat, 4 ggm.fit, 4 \*Topic package XMRF-package, 2 brcadat, 4 ggm.fit, 4 GMS, 4, 5, 5, 7, 11 lambdaMax, 6 plot (plot.GMS), 6 plot.GMS, 5, 6, 11 plotGML, 7 }

plotNet,8 processSeq,9

XMRF, 3, 5, 10 XMRF-package, 2 XMRF.Sim, 12