

# Package ‘DrImpute’

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**Version** 1.0

**Date** 2017-7-15

**Title** Imputing Dropout Events in Single-Cell RNA-Sequencing Data

**Description** R codes for imputing dropout events. Many statistical methods in cell type identification, visualization and lineage reconstruction do not account for dropout events ('PCAreduce', 'SC3', 'PCA', 't-SNE', 'Monocle', 'TSCAN', etc). 'DrImpute' can improve the performance of such software by imputing dropout events.

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**Depends** R (>= 3.1.0)

**Imports** Rcpp

**Suggests** knitr, rmarkdown, devtools, roxygen2, irlba

**License** GPL-3

**VignetteBuilder** knitr

**URL** <https://github.com/ikwak2/DrImpute>

**LinkingTo** Rcpp, RcppArmadillo

**RoxygenNote** 6.0.1

**NeedsCompilation** yes

**Repository** CRAN

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 DrImpute

*Imputing dropout events in single-cell RNA-sequencing data.*


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**Description**

Imputing dropout events in single-cell RNA-sequencing data.

**Usage**

```
DrImpute(X, ks = 10:15, dists = c("spearman", "pearson"), method = "mean",
  cls = NULL, seed = 1, zerop = 0)
```

**Arguments**

|        |  |
|--------|--|
| X      | Gene expression matrix (gene by cell).   |
| ks     | Number of cell clustering groups. Default set to ks = 10:15.   |
| dists  | Distribution matrices to use. Default is set to c("spearman", "pearson"). "euclidean" can be added as well.  |
| method | Use "mean" for mean imputation, "med" for median imputation.   |
| cls    | User can manually provide clustering information. Using different base clusterings. each row represent different clusterings. each column represent each cell. |
| seed   | User can provide a seed.   |
| zerop  | zero percentage of resulting imputation is at least zerop.   |

**Value**

Imputed Gene expression matrix (gene by cell).

**Author(s)**

Il-Youp Kwak

**References**

Il-Youp Kwak, Wuming Gong, Kaoko Koyano-Nakagawa and Daniel J. Garry (2017+) DrImpute: Imputing dropout events in single cell RNA sequencing data

**Examples**

```
data(exdata)
exdata <- preprocessSC(exdata)
exdata <- exdata[1:3000, 1:80]
logdat <- log(exdata+1)
cls <- getCls(logdat)
logdat_imp <- DrImpute(logdat, cls = cls)
```

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 exdata

*Usoskin data*


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### Description

This data set is subset from Usoskin et al. (2015). Original data is RNA-seq data on 799 cells dissected from the mouse lumbar dorsal root ganglion distributed over a total of nine 96-well plates. We randomly selected 150 cells from the data.

Column names indicate four different cell types, NF, NP, TH, and PEP.

### Usage

```
data(exdata)
```

### References

Usoskin D et al. Unbiased classification of sensory neuron types by large-scale single-cell RNA sequencing. Nature Neuroscience. Nature Research,2015;18:145-53.

### Examples

```
data(exdata)
exdata <- preprocessSC(exdata)
```

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 getCls

*get base clustering results using SC3 based clustering methods.*


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### Description

Similarity matrix constructed using "pearson", "spearman" or "euclidean". K-means clustering is performed on first few number of principal components of similarity matrix.

### Usage

```
getCls(X, ks = 10:15, dists = c("spearman", "pearson"),
       dim.reduc.prop = 0.05)
```

### Arguments

|                |   |
|----------------|---|
| X              | Log transformed gene expression matrix (Gene by Cell).  |
| ks             | Number of cell clustering groups. Default set to ks = 10:15.  |
| dists          | Distribution matrices to use. Default is set to c("spearman", "pearson"). "euclidean" can be added as well. |
| dim.reduc.prop | Proportion of principal components to use for K-means clustering.   |

**Value**

A matrix object, Each row represent different clustering results.

**Author(s)**

Il-Youp Kwak

**References**

Il-Youp Kwak, Wuming Gong, Kaoko Koyano-Nakagawa and Daniel J. Garry (2017+) DrImpute: Imputing dropout events in single cell RNA sequencing data

**See Also**

[DrImpute preprocessSC](#)

**Examples**

```
data(exdata)
exdata <- preprocessSC(exdata)
exdata <- exdata[1:3000, 1:80]
logdat <- log(exdata+1)
cls <- getCls(logdat)
```

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preprocessSC

*A function for preprocessing gene expression matrix.*

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**Description**

Preprocess gene expression data

**Usage**

```
preprocessSC(X, min.expressed.gene = 0, min.expressed.cell = 2,
             max.expressed.ratio = 1, normalize.by.size.effect = FALSE)
```

**Arguments**

**X** Gene expression matrix (Gene by Cell).

**min.expressed.gene** Cell level filtering criteria. For a given cell, if the number of expressed genes are less than min.expressed.gene, we filter it out.

**min.expressed.cell** Gene level filtering criteria. For a given gene, if the number of expressed cells are less than min.expressed.cell, we filter it out.

`max.expressed.ratio`

Gene level filtering criteria. For a given gene, if the ratio of expressed cells are larger than `max.expressed.ratio`, we filter it out.

`normalize.by.size.effect`

Normaize using size factor.

### **Value**

Filtered gene expression matrix

### **Author(s)**

Wuming Gong

### **References**

Il-Youp Kwak, Wuming Gong, Kaoko Koyano-Nakagawa and Daniel J. Garry (2017+) DrImpute: Imputing dropout events in single cell RNA sequencing data

### **See Also**

[DrImpute](#)

### **Examples**

```
data(exdata)
exdata <- preprocessSC(exdata)
```

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