# Package 'ACSNMineR'

September 1, 2016

Title Gene Enrichment Analysis from ACSN Maps or GMT Files

Description Compute and represent gene set enrichment or depletion from your

Type Package

**Version** 0.16.8.25

| data based on pre-saved maps from the Atlas of Cancer Signalling Networks (ACSN) or user imported maps. User imported maps must be complying with the GMT format as defined by the Broad Institute, that is to say that the file should be tabseparated, that the first column should contain the module name, the second column can contain comments that will be overwritten with the number of genes in the module, and subsequent columns must contain the list of genes (HUGO |
|--|
| symbols; tab-separated) inside the module. The gene set enrichment can be run with hypergeometric test or Fisher exact test, and can use multiple corrections. Visualization of data can be done either by barplots or heatmaps.   |
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ACSN\_maps

Atlas of Cancer Signalling Networks

# Description

A dataset containing the six maps of ACSN: apoptosis, cell cycle, DNA reparation, EMT motility, survival, and the master map

## Usage

ACSN\_maps

#### **Format**

A list of dataframes

Apoptosis Map of apoptosis pathways

CellCycle Map of the cell cycle pathways

DNA\_repair Map of DNA repair

EMT\_motility Map of the Epithelial Mesenchymal Transition

Master Map grouping all modules from other maps, without a master module for each map

Survival Map of cellular survival pathways

#### Source

https://acsn.curie.fr/downloads.html

cnum 3

cnum

Convert to numeric

# Description

Convert to numeric

# Usage

cnum(x)

## **Arguments**

Х

A vector of numbers which is not in numeric format

Create\_master\_map

From a list of maps, create or replace a master

# Description

From a list of maps, create or replace a master

## Usage

```
Create_master_map(maps)
```

## **Arguments**

maps

A list of molecular maps created by 'format\_from\_gmt'

## Value

Returns a list with previous maps and the master map, i.e. a concatenation of previous maps.

# **Examples**

4 enrichment

enrichment

Gene set enrichment analysis

# Description

Compute and represent gene set enrichment from your data based on pre-saved maps from ACSN or user imported maps. The gene set enrichment can be run with hypergeometric test or Fisher exact test, and can use multiple corrections. Visualization of data can be done either by barplots or heatmaps.

## Usage

```
enrichment(Genes = NULL, maps = c("Apoptosis", "CellCycle", "DNA_repair",
   "EMT_motility", "Survival"), correction_multitest = "BH",
   statistical_test = "fisher", min_module_size = 5,
   universe = "map_defined", Remove_from_universe = NULL, threshold = 0.05,
   alternative = "greater")
```

#### **Arguments**

Genes Character vector of genes that should be tested for enrichment

maps list of maps generated by format\_from\_gmt. Names of element of list will be

used to track modules. Default: tests on the master map.

correction\_multitest

either FALSE, "bonferroni", "holm", "hochberg", "hommel", "BH", "fdr" (iden-

tical to BH), or "BY"

statistical\_test

one of "fisher", "hypergeom"

min\_module\_size

will remove from the analysis all modules which are (strictly) smaller than

threshold

universe Universe on which the statistical analysis should be performed. Can be either

"HUGO", "ACSN", "map\_defined", or a character vector of genes.

Remove\_from\_universe

Default is NULL. A list of genes that should not be considered for enrichment

(will be removed from input, maps, and universe). The size of universe and map

will be updated after removal.

threshold maximal p-value (corrected if correction is enabled) that will be displayed

alternative One of "greater", "less", "both" or "two.sided" Greater will check for enrich-

ment, less will check for depletion, and both will look for both and will keep track of the side, while two-sided (only for fisher test) checks if there is a differ-

ence.

enrichment\_test 5

#### Value

Output is a dataframe with the following columns:

module The name of the map or the module preceded by the map
module\_size The number of genes in the module after taking into account universe reduction
nb\_genes\_in\_module The number of genes from input list in the module
genes\_in\_module Names of the genes from input list in the module, space separated
universe\_size size of the input universe
nb\_genes\_in\_universe number of genes from the input list that are found in the universe
test the kind of test that was looked for. "greater" when enrichment is tested, "less" when depletion is tested, or "two.sided"

## **Examples**

```
enrichment(genes_test,min_module_size = 10,
    threshold = 0.05,
    maps = list(cellcycle = ACSNMineR::ACSN_maps$CellCycle),
    universe = "ACSN")
```

enrichment\_test

Result from enrichment test of "genes\_test" on the ACSN maps

#### **Description**

Parameters: bonferroni correction, min module size = 5

## Usage

```
enrichment_test
```

#### Format

data.frame

**module** Name of module

genes\_in\_module Genes from genes\_test in module

p.value Uncorrected p-value

p.value.corrected p-value corrected for multiple testing by Bonferroni correction

genes\_test

format\_from\_gmt

Import data from gmt files Convert gmt file to dataframe that can be used for anaysis

## **Description**

Import data from gmt files Convert gmt file to dataframe that can be used for anaysis

# Usage

```
format_from_gmt(path = "")
```

## Arguments

path

Path to the gmt file to be imported

## Value

Returns a dataframe with the module - first column -, module length - seconde column - and gene names

# **Examples**

```
file<-system.file("extdata", "cellcycle_short.gmt", package = "ACSNMineR")
format_from_gmt(file)</pre>
```

genes\_test

Set of genes to test map

# Description

Genes of high importance in oncogenesis

## Usage

```
{\tt genes\_test}
```

#### **Format**

A character vector

```
multisample_enrichment
```

Automated gene set analysis for multiple sets

#### **Description**

Automated gene set analysis for multiple sets

#### Usage

```
multisample_enrichment(Genes_by_sample = NULL, maps = c("Apoptosis",
   "CellCycle", "DNA_repair", "EMT_motility", "Survival"),
   correction_multitest = "BH", statistical_test = "fisher",
   min_module_size = 5, universe = "map_defined",
   Remove_from_universe = NULL, threshold = 0.05, cohort_threshold = TRUE,
   alternative = "greater")
```

#### Arguments

Genes\_by\_sample

List of character vectors. Each list element name should be a sample name, and each character vector the set of genes to test for the sample.

maps list of maps generated by format\_from\_gmt. Default: tests on all acsn maps correction multitest

either FALSE, "bonferroni", "holm", "hochberg", "hommel", "BH", "fdr" (identical to BH), or "BY"

statistical\_test

one of "fisher", "hypergeom"

min\_module\_size

will remove from the analysis all modules which are (strictly) smaller than threshold

universe

Universe on which the statistical analysis should be performed. Can be either "HUGO", "ACSN", "map\_defined", or a character vector of genes.

Remove\_from\_universe

pendently.

Default is NULL. A list of genes that should not be considered for enrichment (will be removed from input, maps, and universe). The size of universe and map will be updated after removal.

threshold rochort\_threshold

maximal p-value (corrected if correction is enabled) that will be displayed

if TRUE modules will be kept in all samples if at least one sample has p-value lower than threshold, otherwise the threshold is applied for each sample inde-

alternative

One of "greater", "less", "both", or "two.sided" (only for fisher test). Greater will check for enrichment, less will check for depletion, and both will look for both.

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#### Value

Output is a list of dataframes with names the names given in 'Genes\_by\_sample' with the following columns:

module The name of the map or the module preceded by the map
module\_size The number of genes in the module after taking into account universe reduction
nb\_genes\_in\_module The number of genes from input list in the module
genes\_in\_module Names of the genes from input list in the module, space separated
universe\_size size of the input universe

**nb\_genes\_in\_universe** number of genes from the input list that are found in the universe **test** the kind of test that was looked for. "greater" when enrichment is tested, "less" when depletion is tested, or "two.sided"

# Examples

```
multisample_enrichment(Genes_by_sample = list(set1 = genes_test,set2=c(genes_test,"PTPRD")),
maps = list(cellcycle = ACSNMineR::ACSN_maps$CellCycle),
min_module_size = 10,
universe = "ACSN",cohort_threshold = FALSE)
```

p.val.calc

Calculate p-value

#### Description

Calculate p-value

#### Usage

```
p.val.calc(x, y, z, a, stat_test, alt)
```

## **Arguments**

x : first valuey : second valuez : third valuea : fourth value

stat\_test : statistical test to be used

alt : alternative: one of two-sided, greater, less or both

represent\_enrichment 9

represent\_enrichment Graphic representation of enrichment

# Description

Graphic representation of enrichment

# Usage

```
represent_enrichment(enrichment, plot = "heatmap", scale = "log",
  low = "steelblue", high = "white", nrow = 1, sample_name = "Sample",
  na.value = "grey")
```

## **Arguments**

| enrichment  | Data frame or list of dataframes with p-values or corrected p-values (whenever available) and module names for representation. The name of the dataframe will be used as sample name. |
|-------------|---|
| plot        | Any of "heatmap" or "bar"   |
| scale       | Any of "log", "identity" or "reverselog" (i.elog10(p-value))  |
| low         | Color to be used in heatmap mode corresponding to lowest value  |
| high        | Color to be used in heatmap mode corresponding to highest value   |
| nrow        | Number of rows of the grid for display in bar mode.   |
| sample_name | used only is enrichment is a dataframe  |
| na.value    | color for the missing values in the heatmap   |

## Value

Function returns a ggplot2 object if input is a dataframe or a gridExtra object if the output is a list.

# **Examples**

10 reverselog\_trans

reverselog\_trans

Scale for barplots and heatmaps

# Description

Outputs the "-log" of a scale

# Usage

```
reverselog_trans(base = 10)
```

# Arguments

base

: base for the log, defaut is 10

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